



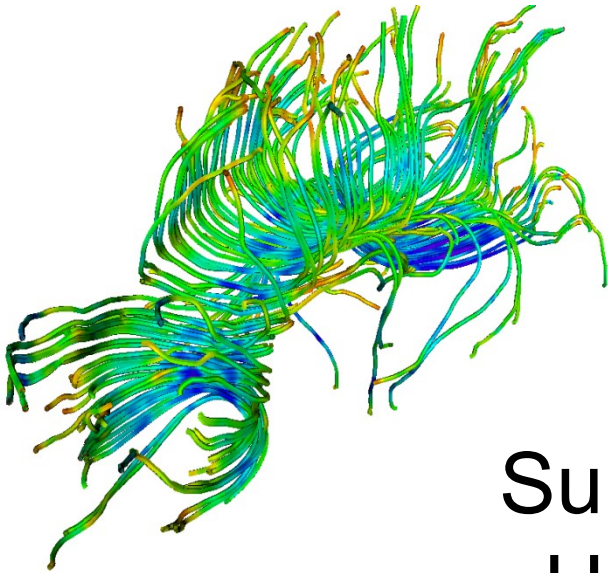
NA-MIC

*National Alliance for Medical Image Computing*

*<http://www.na-mic.org>*

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# Diffusion Tensor Imaging Tutorial



Sonia Pujol, Ph.D.

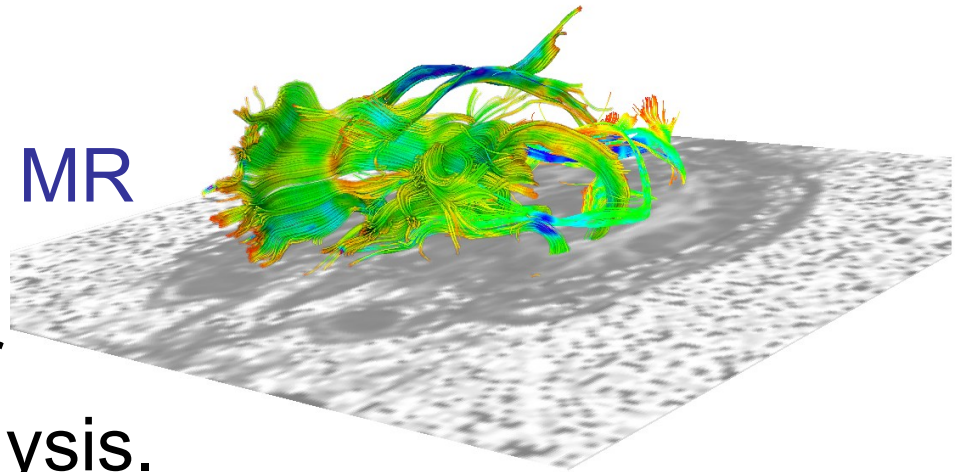
Surgical Planning Laboratory  
Harvard Medical School



# DTI tutorial

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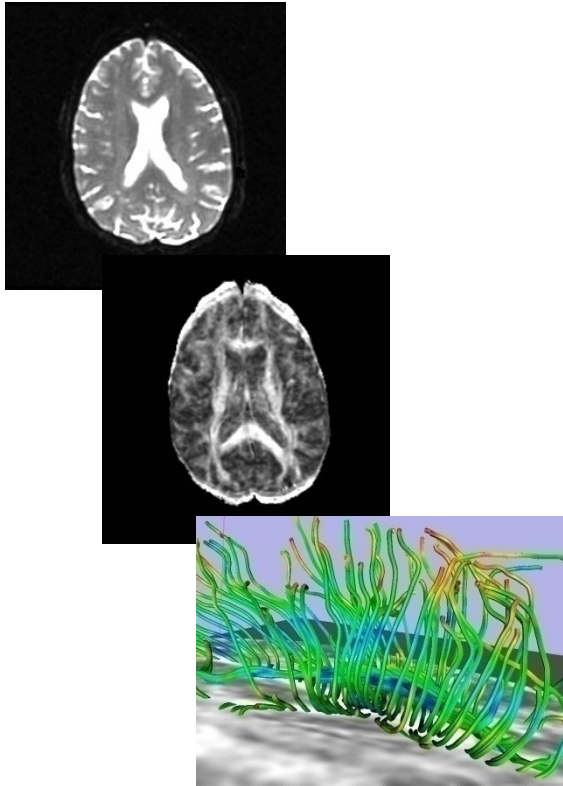
This tutorial is an introduction to the advanced **Diffusion MR** capabilities of the **Slicer3** software for medical image analysis.





# Outline

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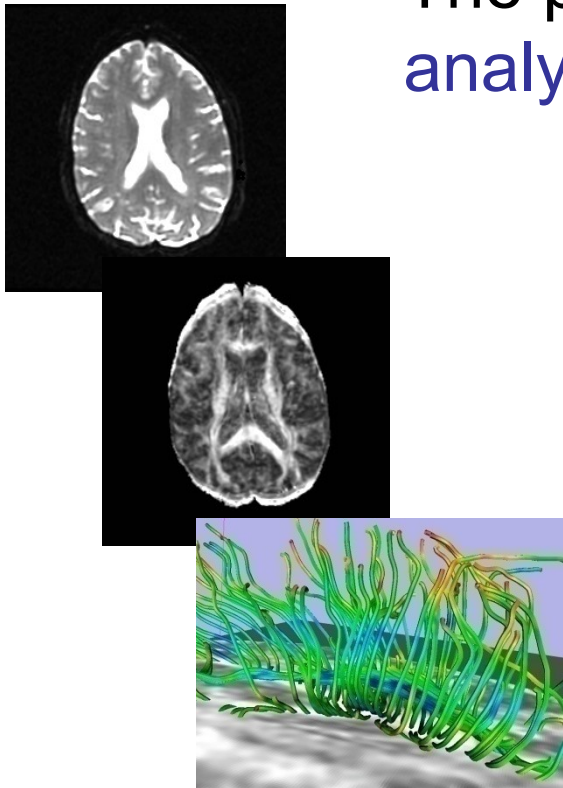
This tutorial guides you through the process of **loading diffusion MR data**, **estimating diffusion tensors**, and performing **tractography** of white matter bundles.



# Outline

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The processing pipeline uses 9 image analysis modules of Slicer3.6



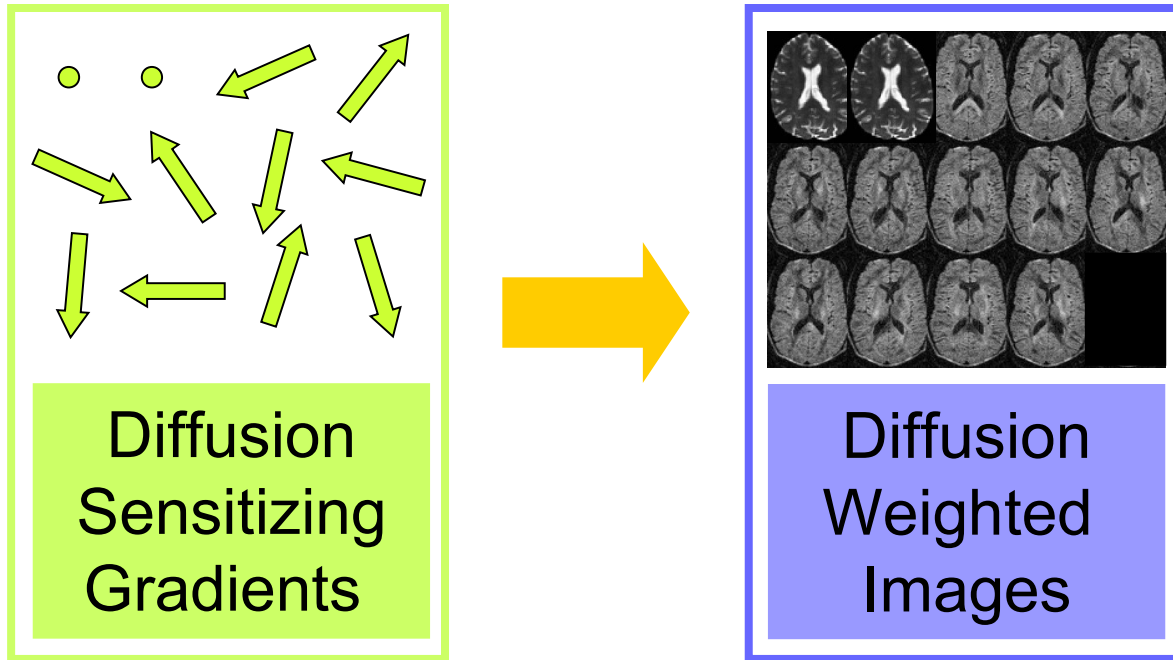
1. Data
2. Volumes
3. Diffusion Tensor Estimation
4. Diffusion Tensor Scalar Measurements
5. Editor
6. LabelMap Seeding
7. Fiber Bundles
8. Fiducials
9. Fiducial Seeding





# Tutorial Dataset

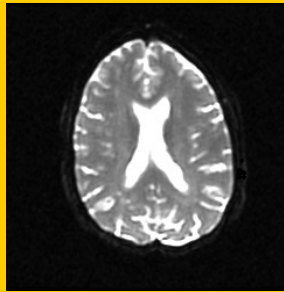
The Diffusion MR tutorial dataset is composed of a **Diffusion Weighted MR scan** of the brain acquired with 12 gradient directions and 2 baseline.



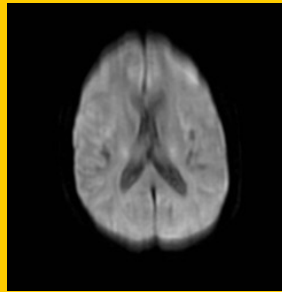


# DTI Processing Pipeline

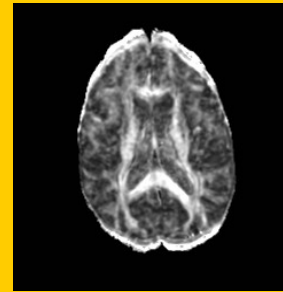
---



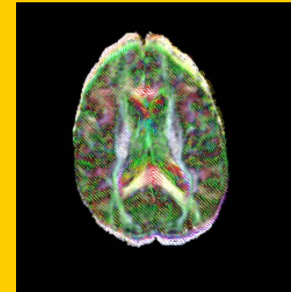
DWI  
Acquisition



Tensor  
Calculation



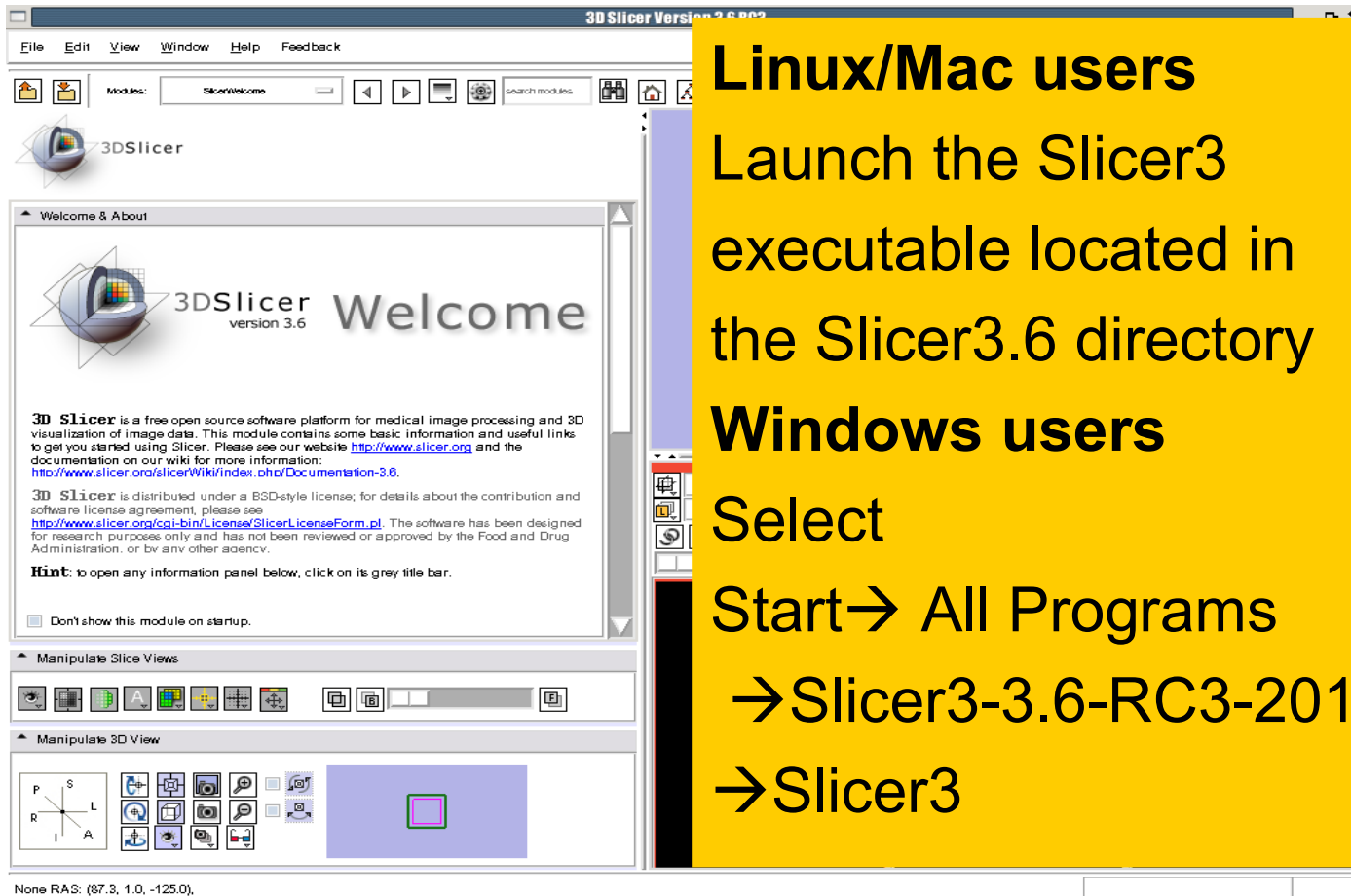
Scalar  
Maps



3D  
Visualization



# Start Slicer3



**Linux/Mac users**

Launch the Slicer3  
executable located in  
the Slicer3.6 directory

**Windows users**

Select

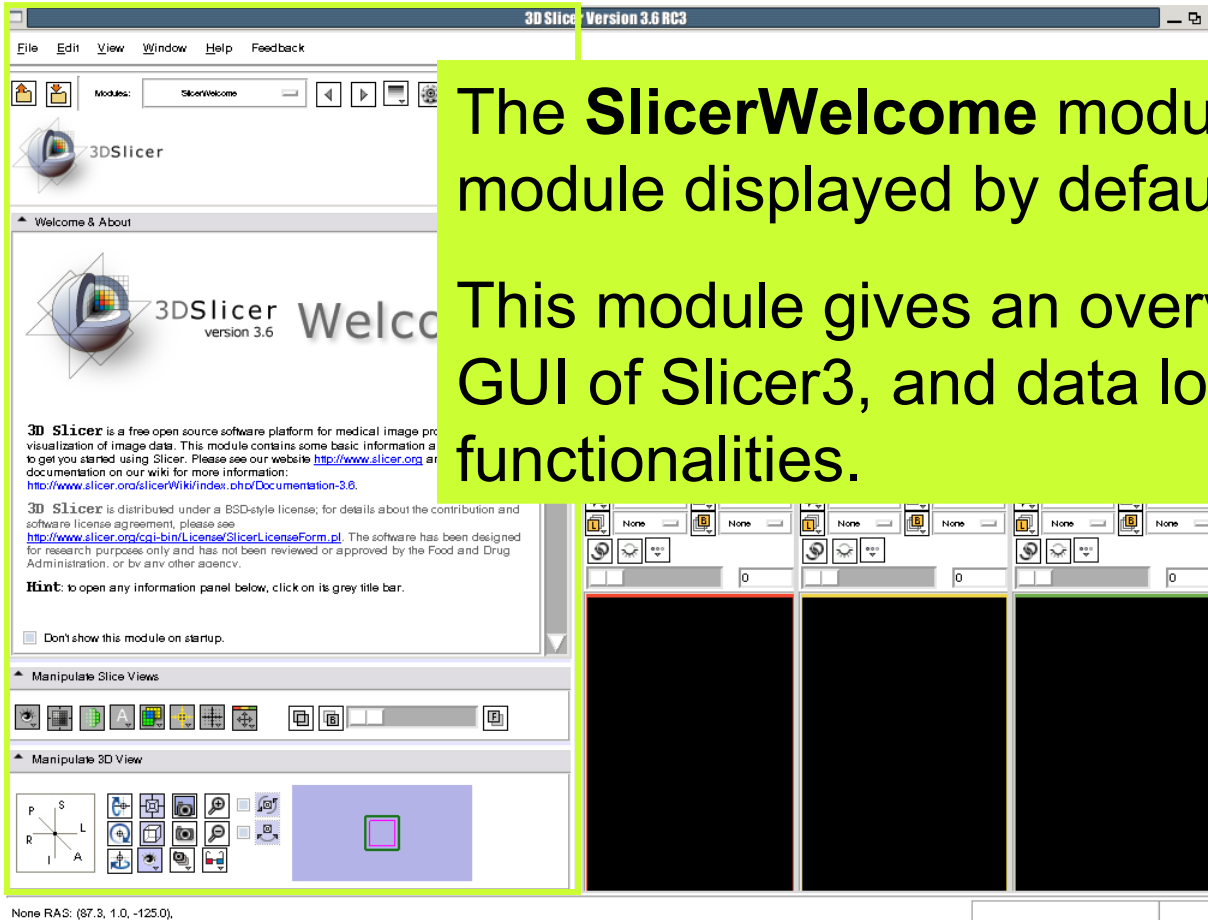
Start → All Programs

→ Slicer3-3.6-RC3-2010-06-04

→ Slicer3



# Slicer Welcome



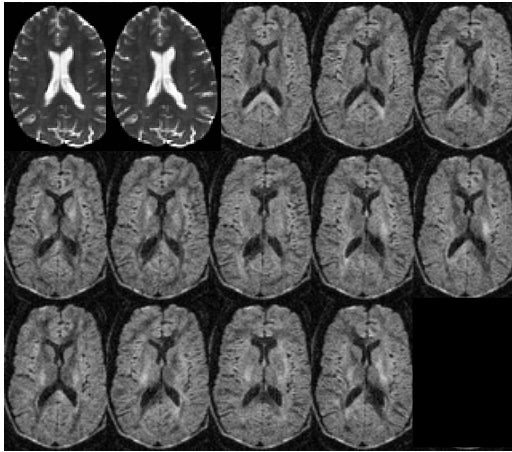
The **SlicerWelcome** module is the module displayed by default.

This module gives an overview of the GUI of Slicer3, and data loading & saving functionalities.



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## Part 1:

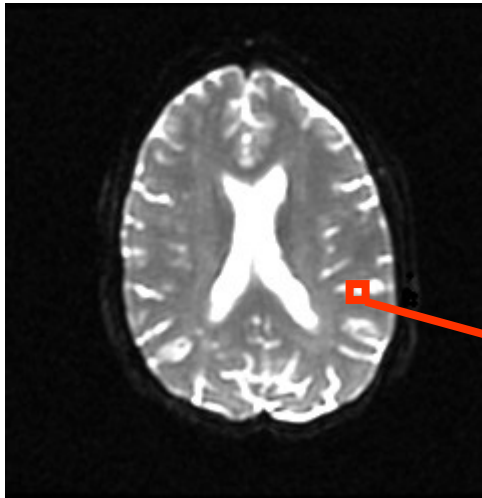


# Diffusion data loading and tensor estimation



# Diffusion Tensor

Stejskal-Tanner



$$S_i = S_0 e^{-b \hat{g}_i^T \underline{D} \hat{g}_i}$$

$$\underline{\mathbf{D}} = \begin{bmatrix} D_{xx} & D_{xy} & D_{xz} \\ D_{yx} & D_{yy} & D_{yz} \\ D_{zx} & D_{zy} & D_{zz} \end{bmatrix}$$



# Tutorial Dataset

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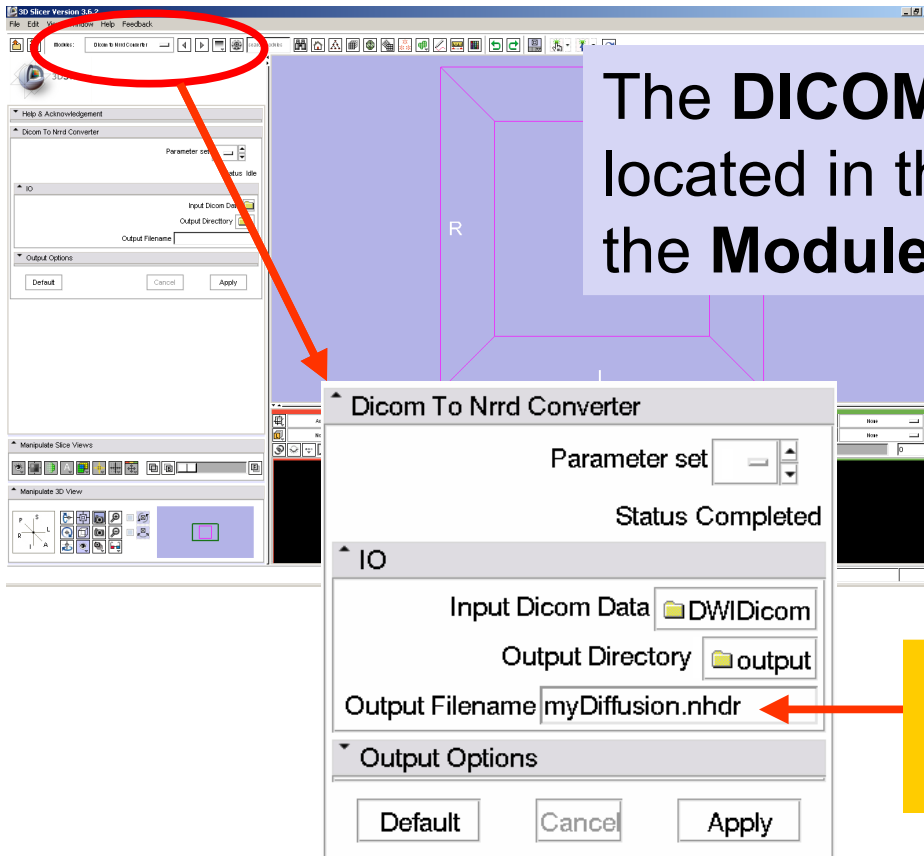
The dataset used in this tutorial is in **the Nrrd file format**, which is part of the NA-MIC kit.

To convert your own Dicom data to Nrrd, use the **DicomToNrrdConverter** module in Slicer.





# DicomToNrrd converter



The **DICOMToNrrdConverter** is located in the **Converters** category in the **Modules** menu

Use **.nhdr** for **Output Filename** extension



# DicomToNrrd converter



search

Google Custom Search

GO

navigation

- Slicer website
- Wiki Home
- Slicer Downloads
- Training
- Documentation
- Users
- Developers
- FAQ
- Acknowledgements
- Links
- Recent Changes

toolbox

- What links here
- Related changes
- Special pages
- Printable version
- Permanent link

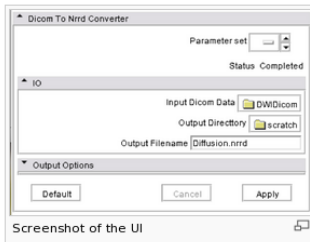
[page](#) [discussion](#) [view source](#) [history](#)

Modules:DicomToNRRD-3.6

[Return to Slicer 3.6 Documentation](#)

Module Name

DWI Dicom To NRRD



Screenshot of the UI

GENERAL INFORMATION

Module Type & Category

Type: Command line module

Category: Converters

Authors, Collaborators & Contact

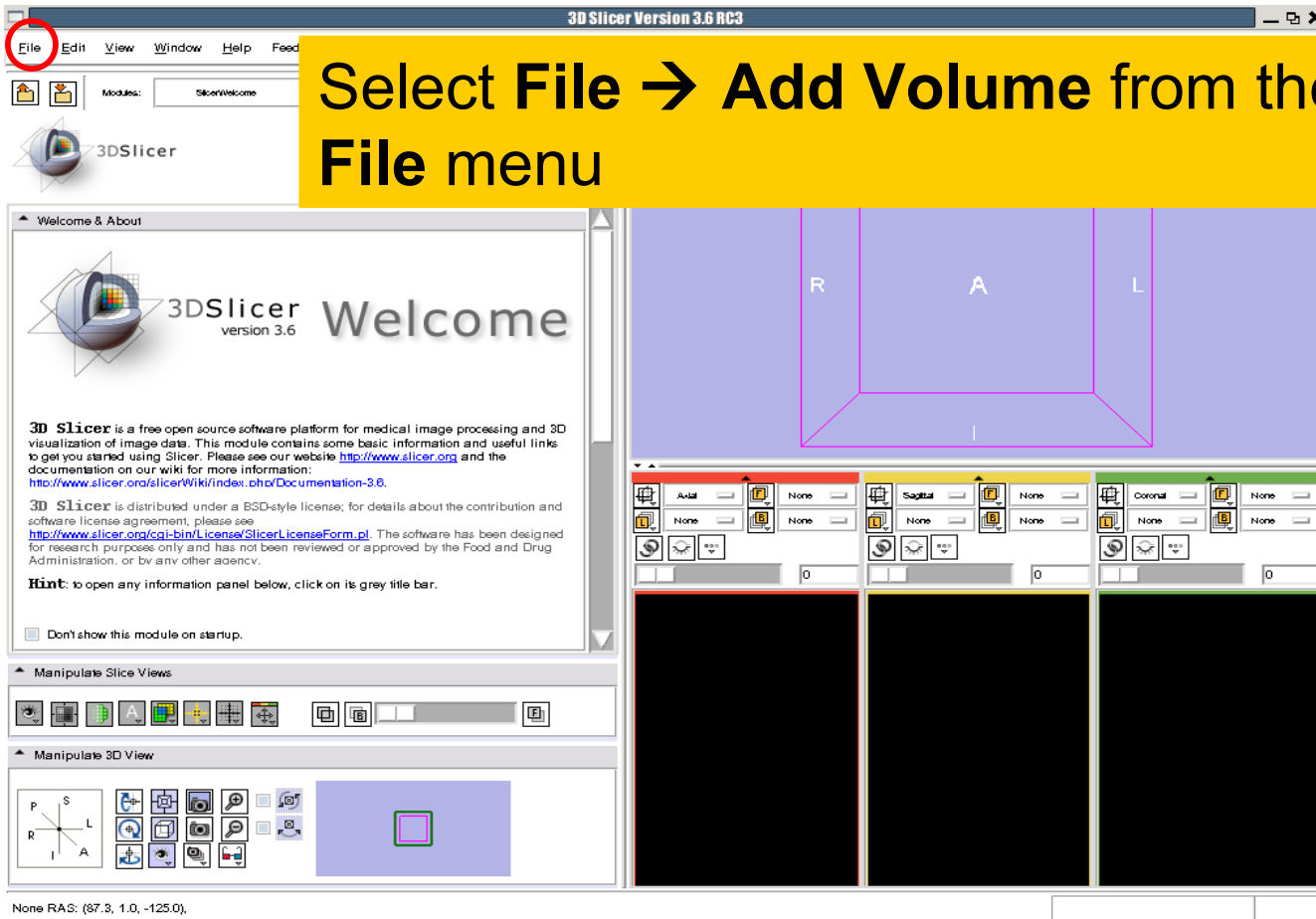
- Author: Xiaodong Tao (with contribution from Vince Magnotta and Hans Johnson)

A list of supported DWI formats can be found on the documentation page of the **DicomToNrrdconverter**:

<http://www.slicer.org/slicerWiki/index.php/Modules:DicomToNRRD-3.6>

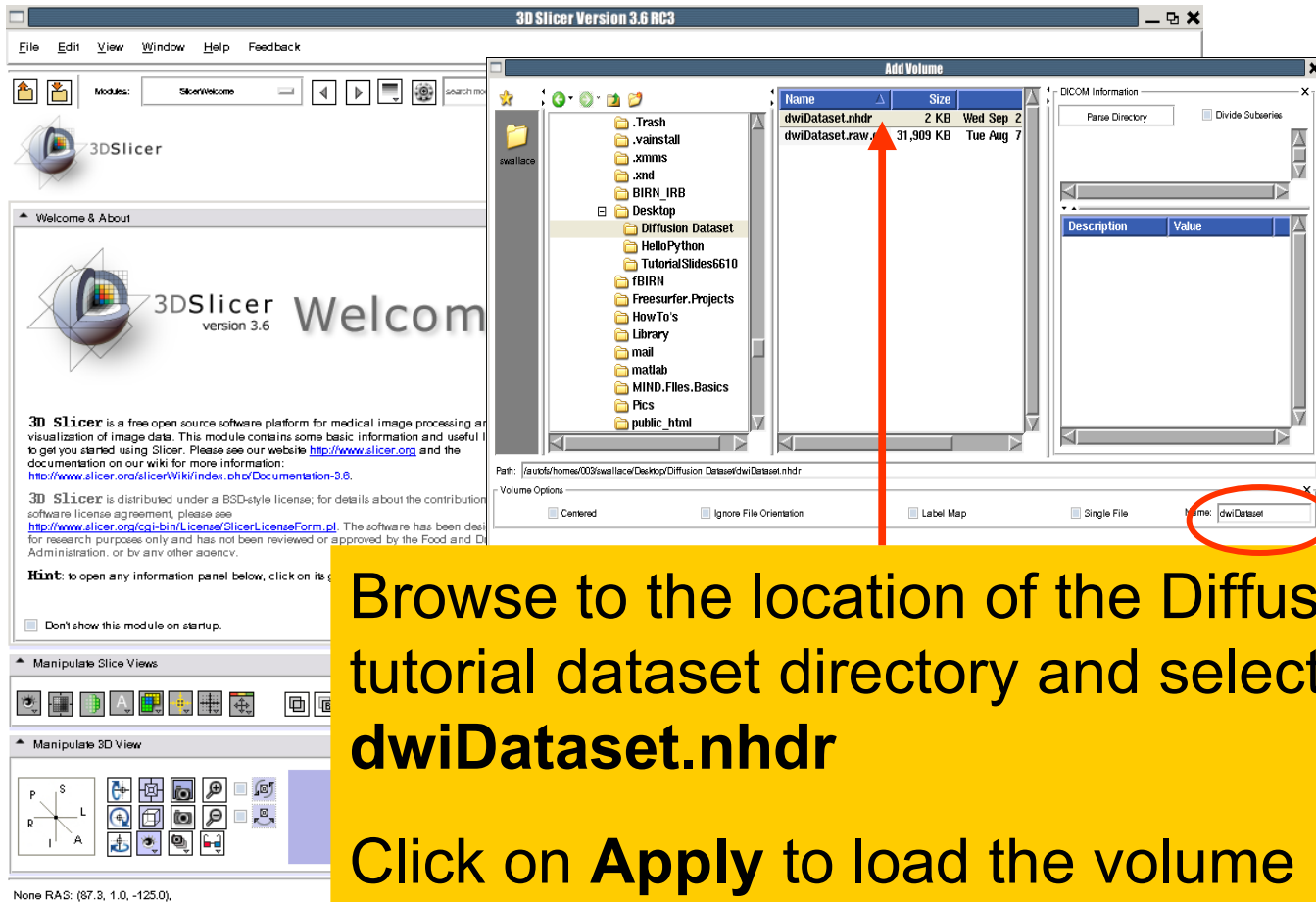


# Loading the DWI Volume





# Loading the DWI Volume

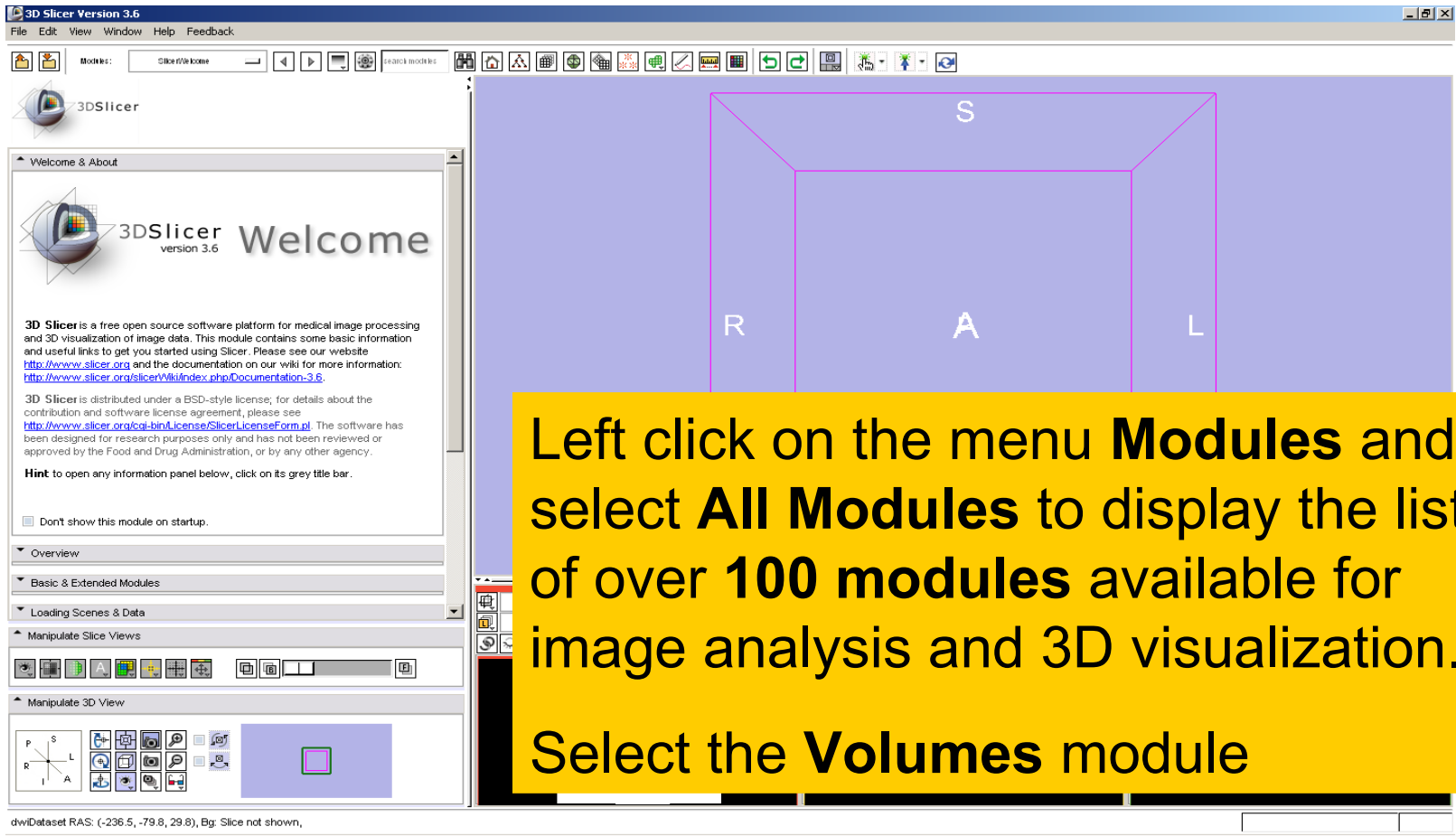


Browse to the location of the Diffusion tutorial dataset directory and select the file **dwiDataset.nhdr**

Click on **Apply** to load the volume



# Loading the DWI Volume





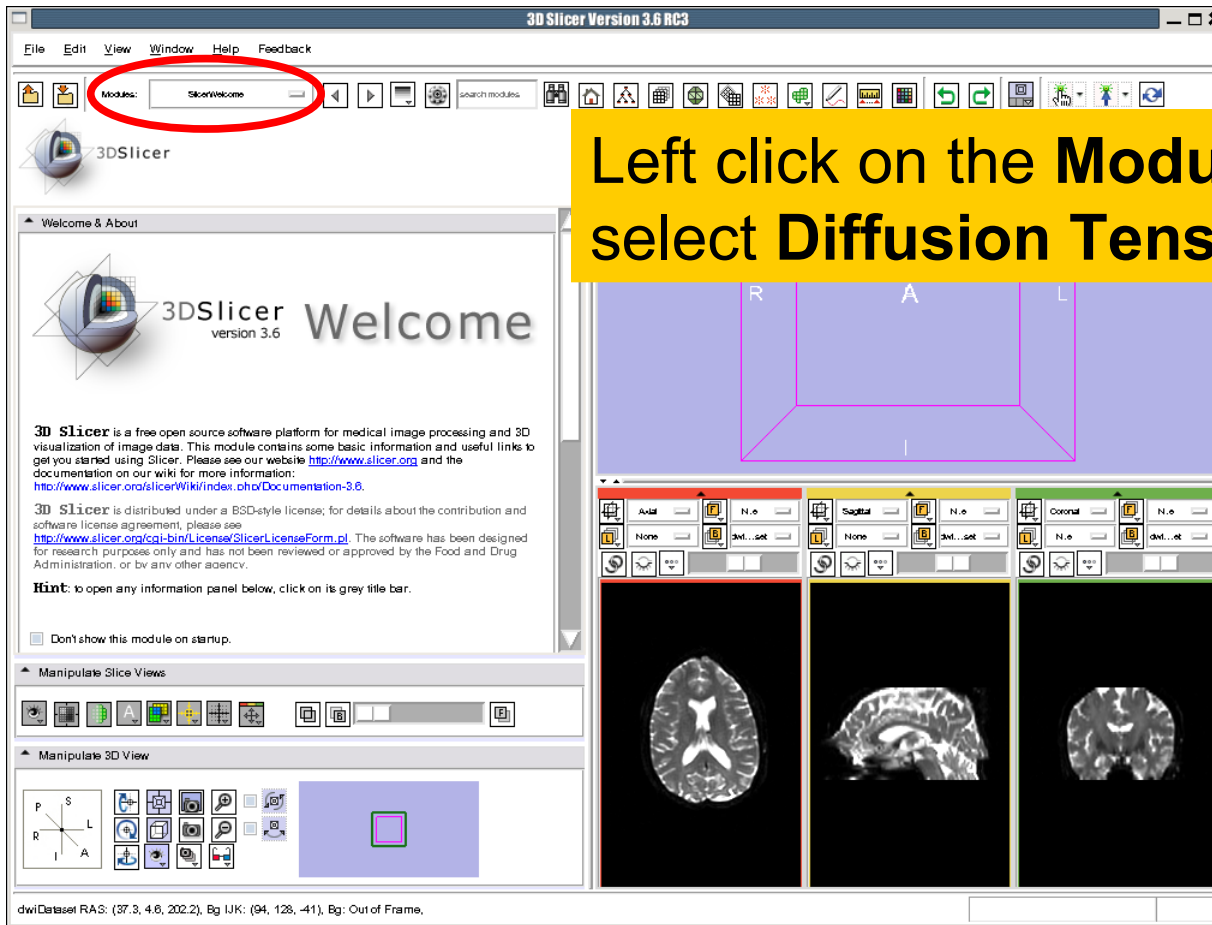
# Loading the DWI Volume

Select the Active Volume **dwiDataset** and adjust the Window/Level Parameters

Slicer displays the anatomical views of the baseline volume of the diffusion dataset in the 2D Slice Viewer.



# Tensor Estimation

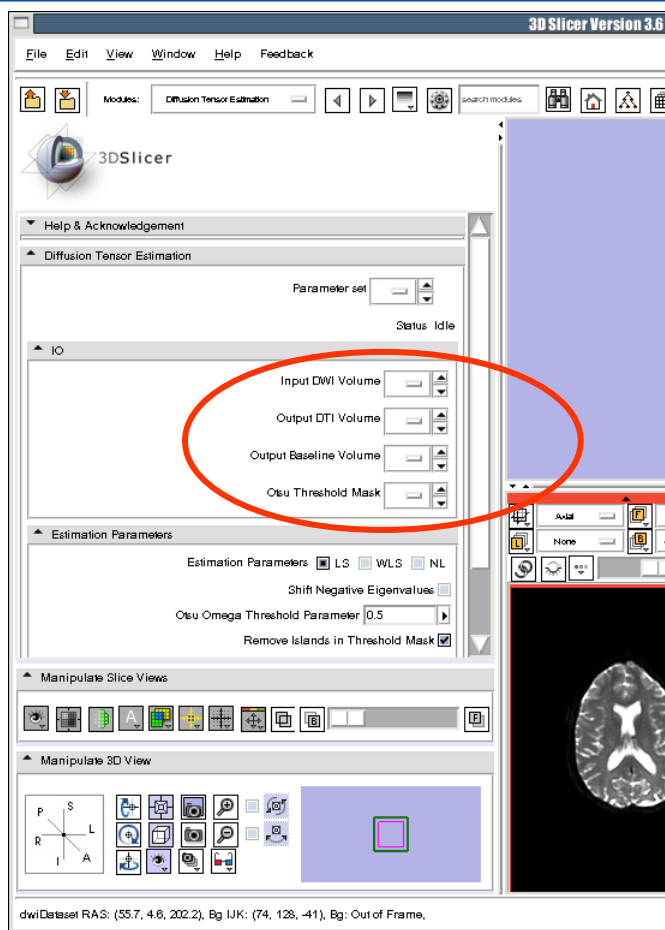


Left click on the **Modules** menu and select **Diffusion Tensor Estimation**.





# Tensor Estimation



Select the Input DWI Volume  
**dwiDataset**

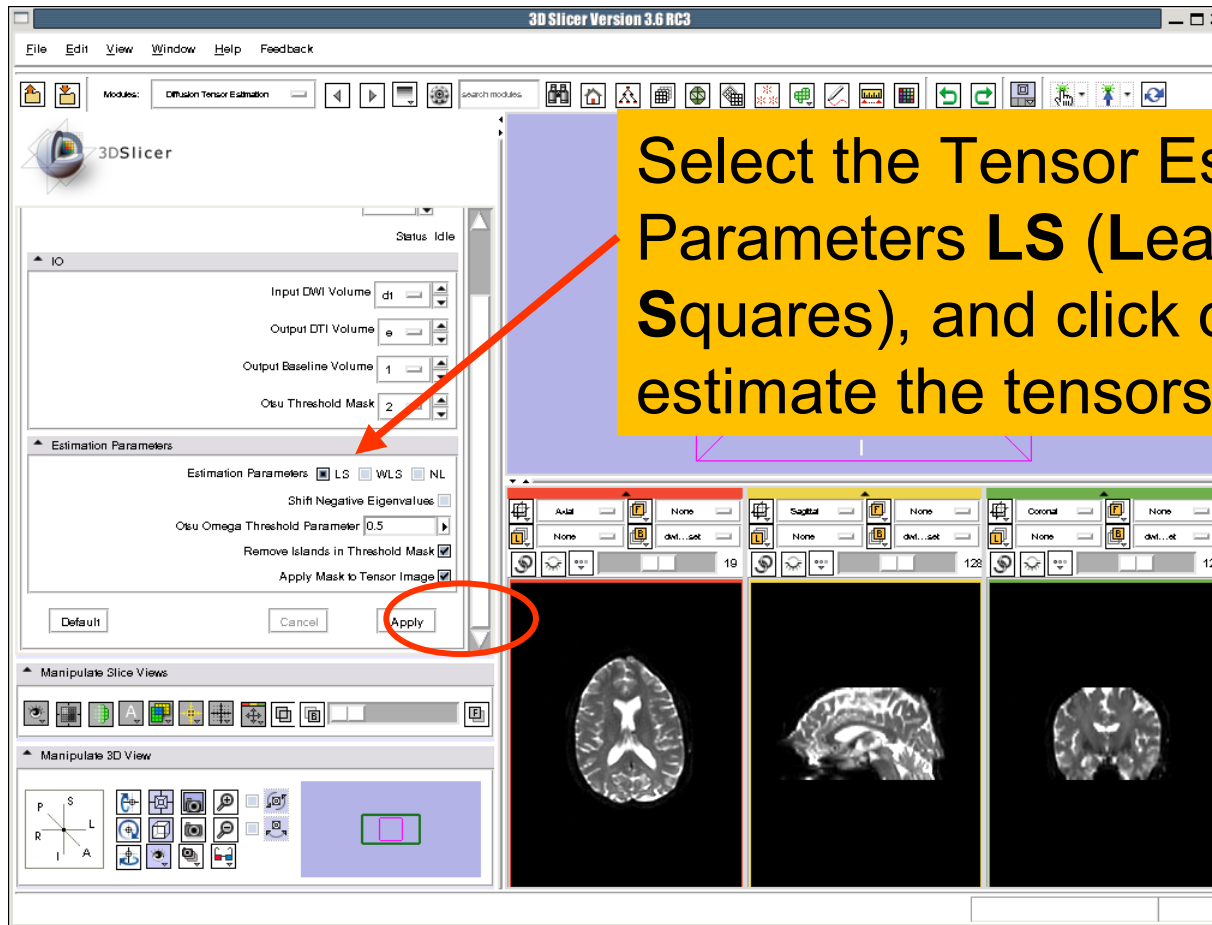
Left click on **OutputDTIVolume** and  
select **Create New Diffusion  
Tensor Volume**

Left click on **Output Baseline  
Volume** and select **Create New  
Volume**

Left click on **Otsu Threshold Mask**  
and select **Create New Volume**



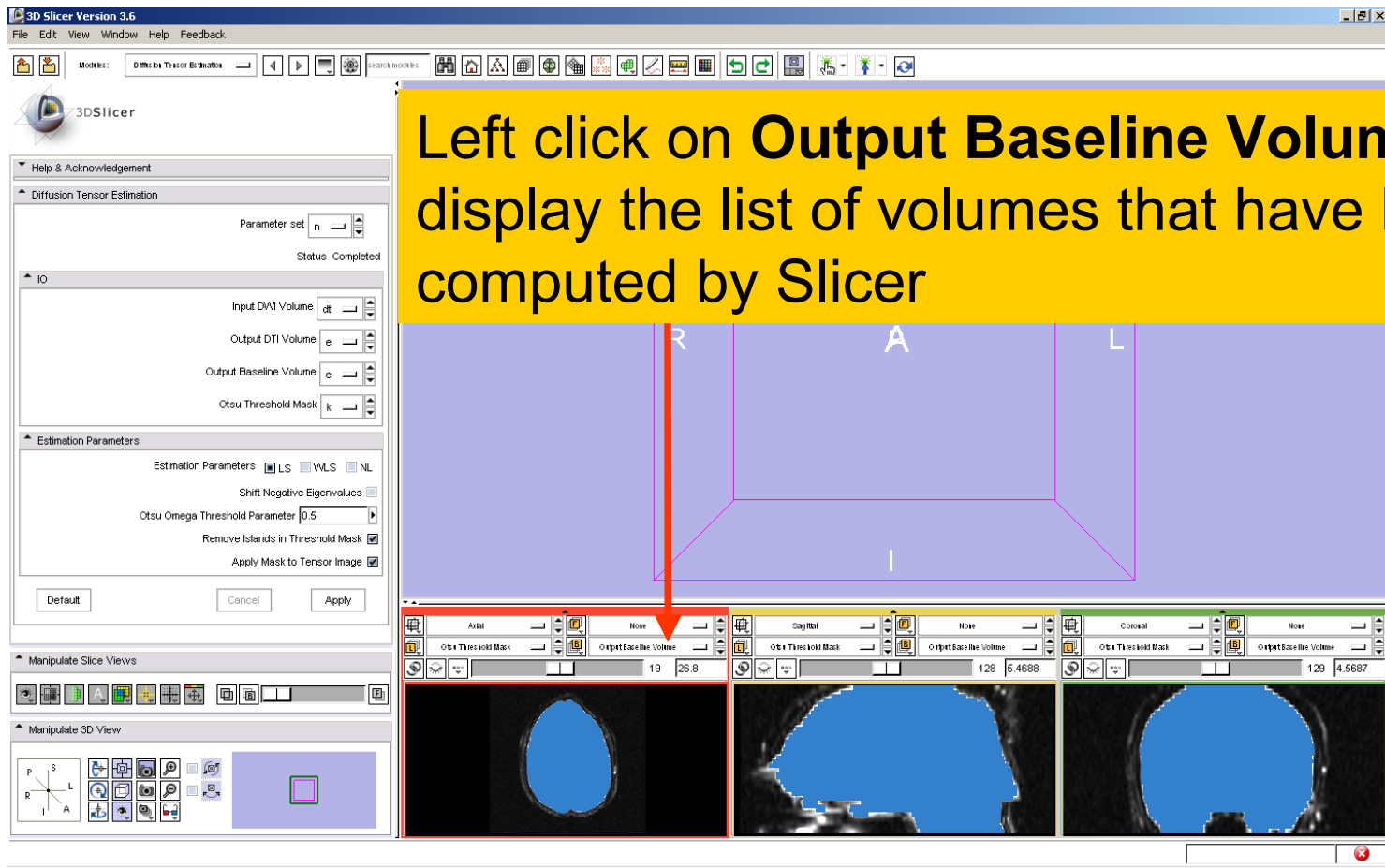
# Tensor Estimation



Select the Tensor Estimation Parameters **LS** (Least Squares), and click on **Apply** to estimate the tensors.



# Tensor Estimation



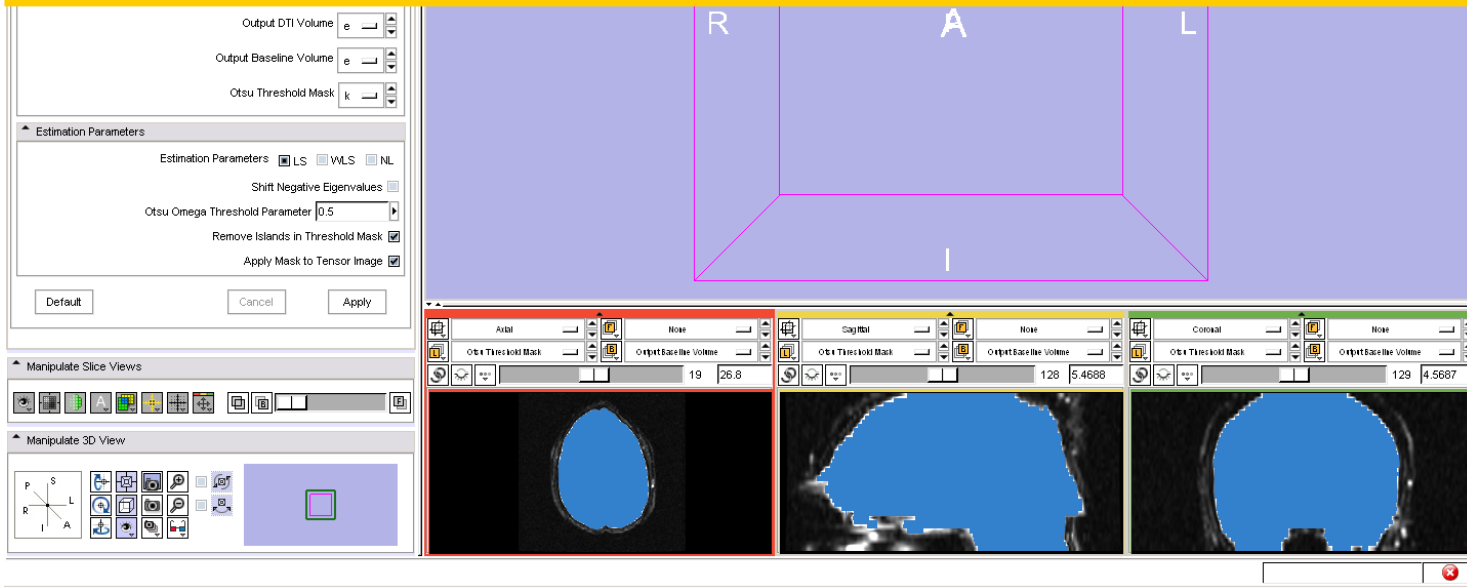


# Tensor Estimation

**Output DTI Volume** is the volume of estimated tensors

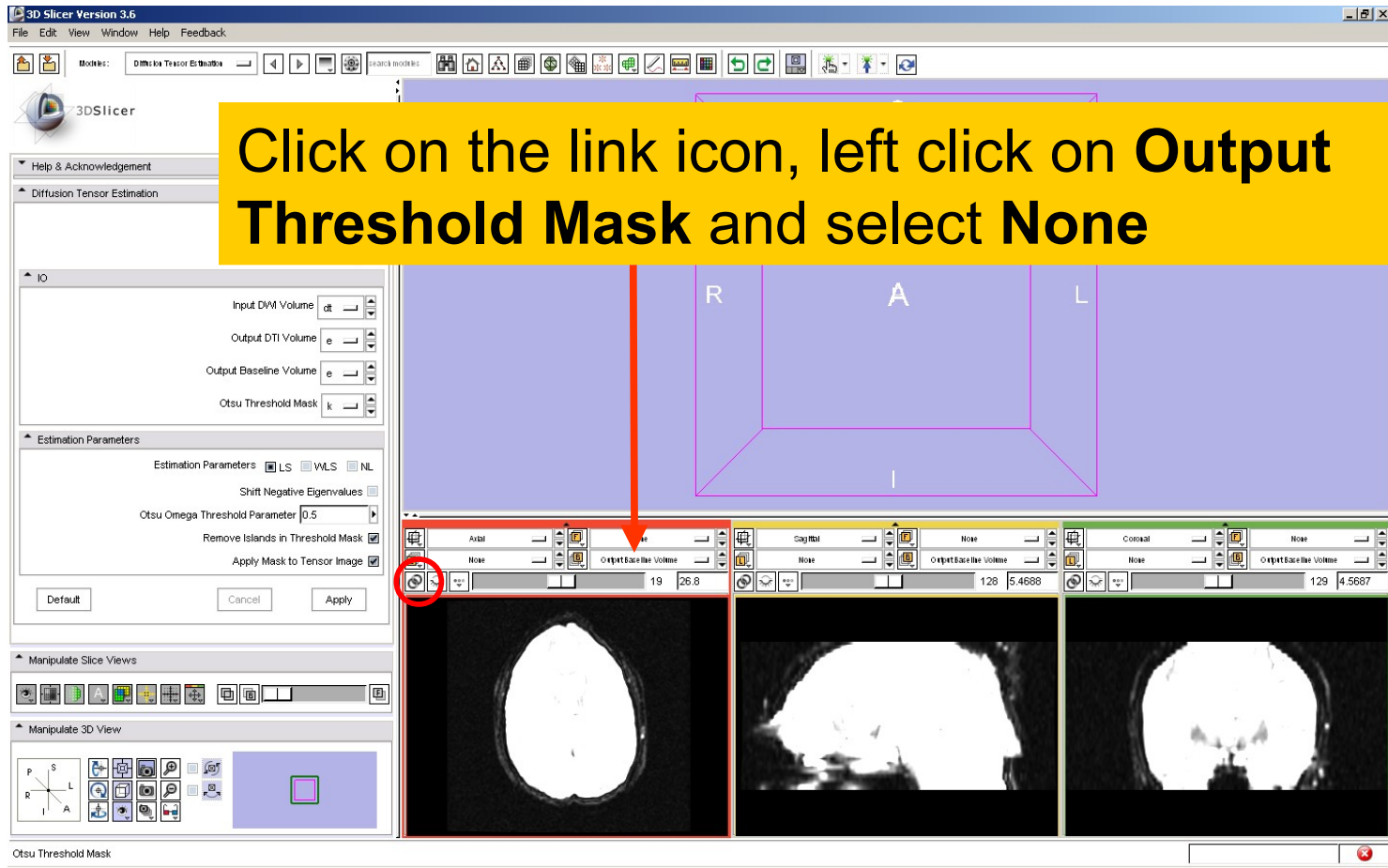
**Output Baseline Volume** is the Baseline volume

**Otsu Threshold Mask** is the tensor mask (blue)



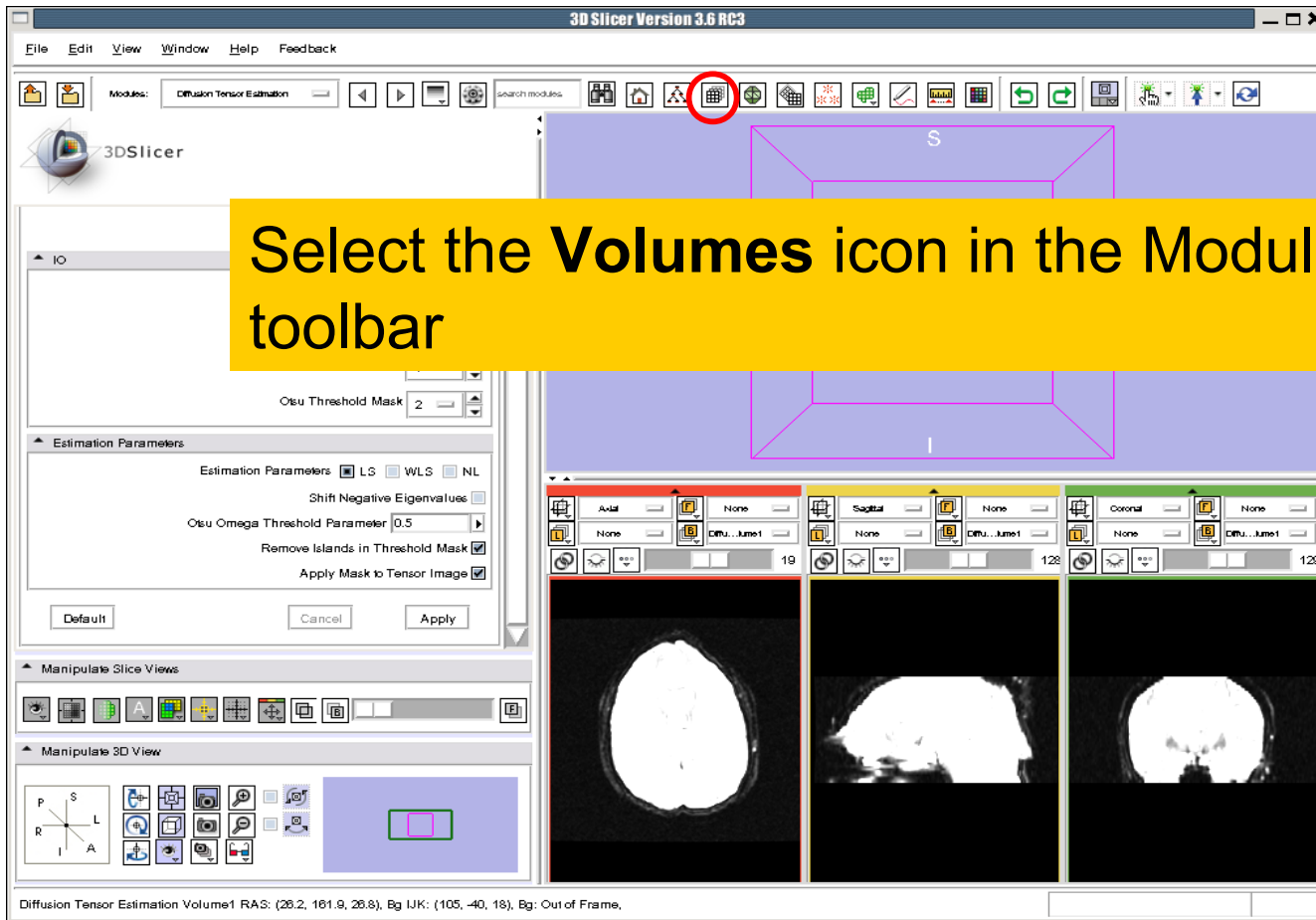


# Tensor Estimation



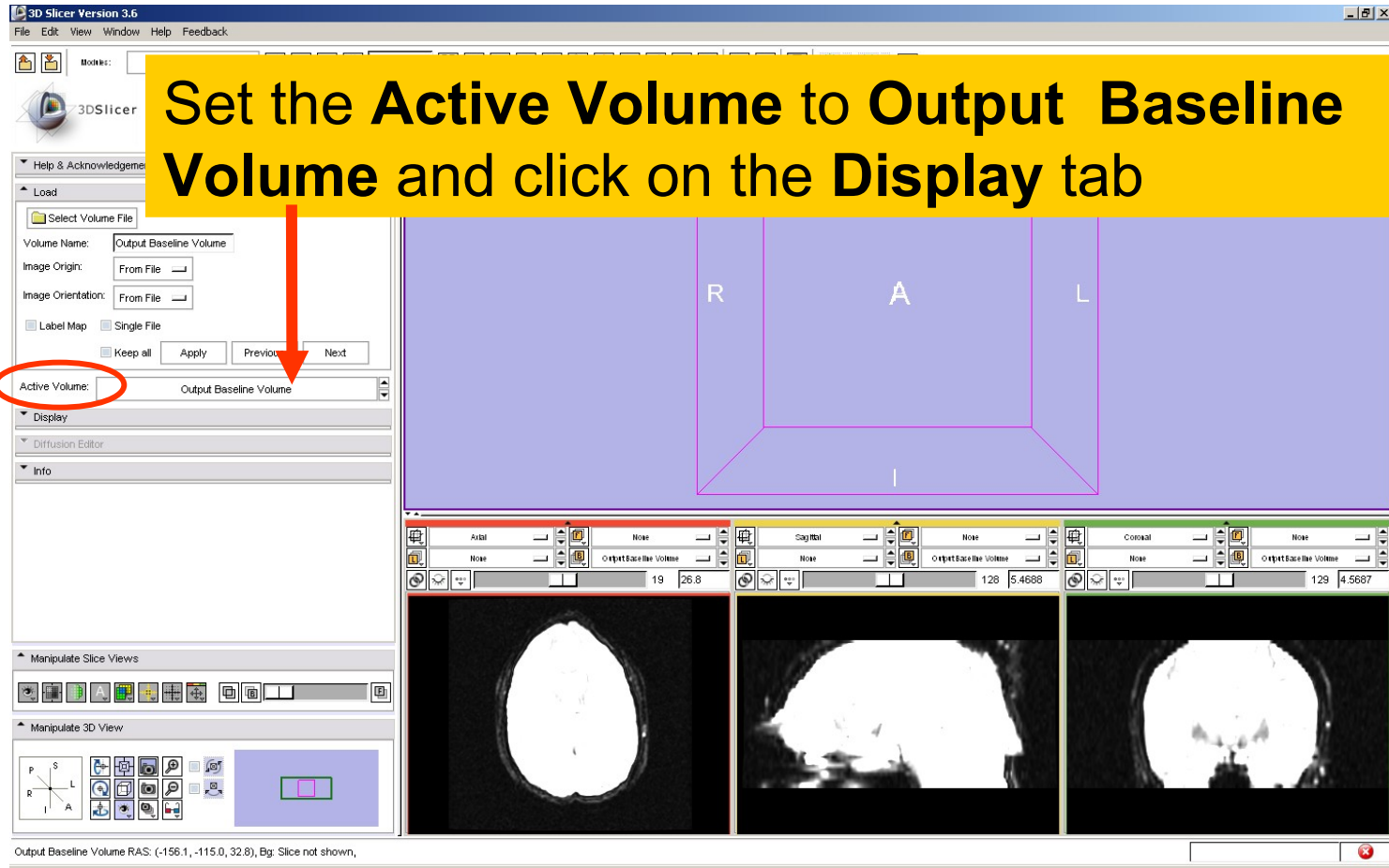


# Tensor Estimation





# Tensor Estimation

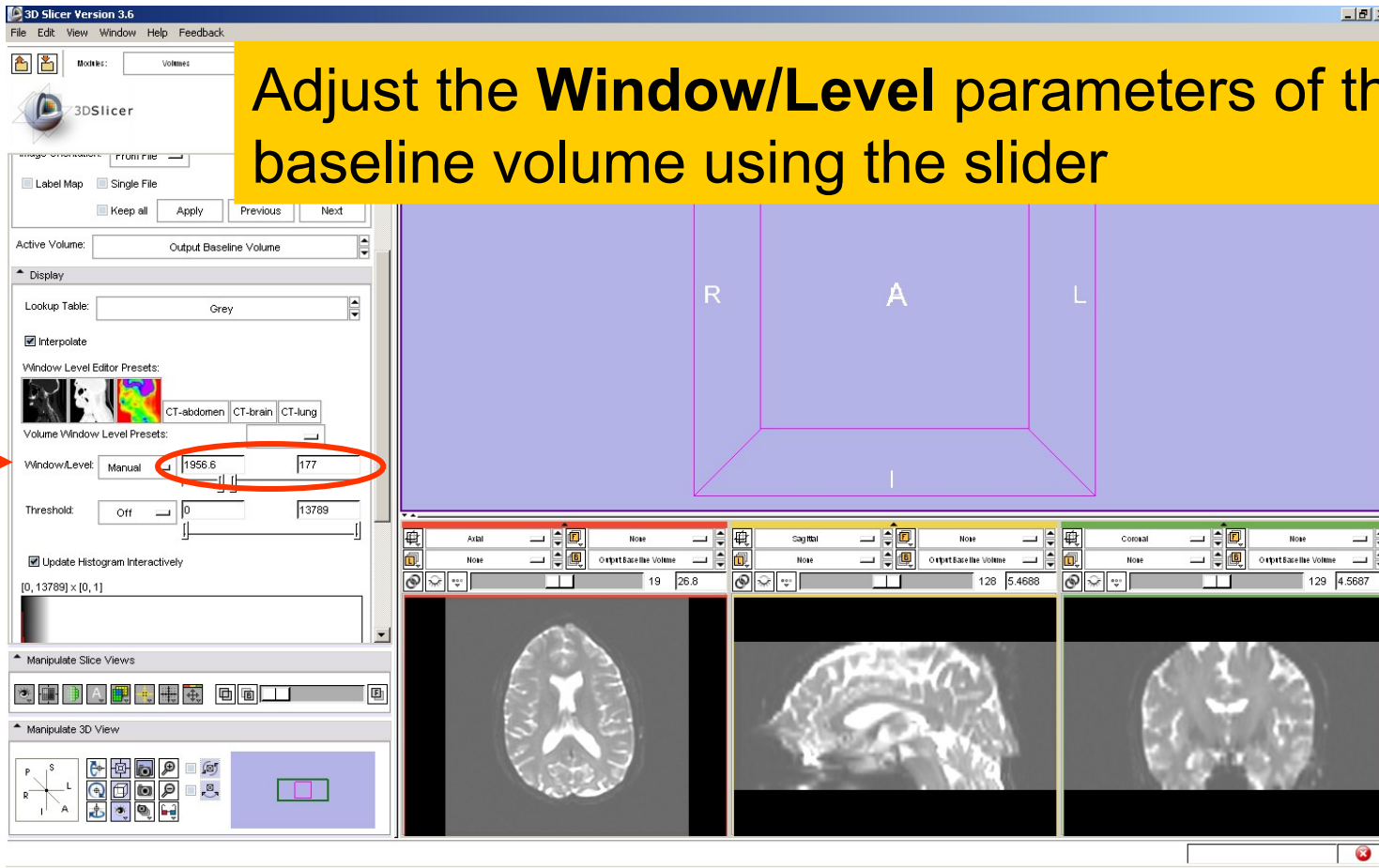






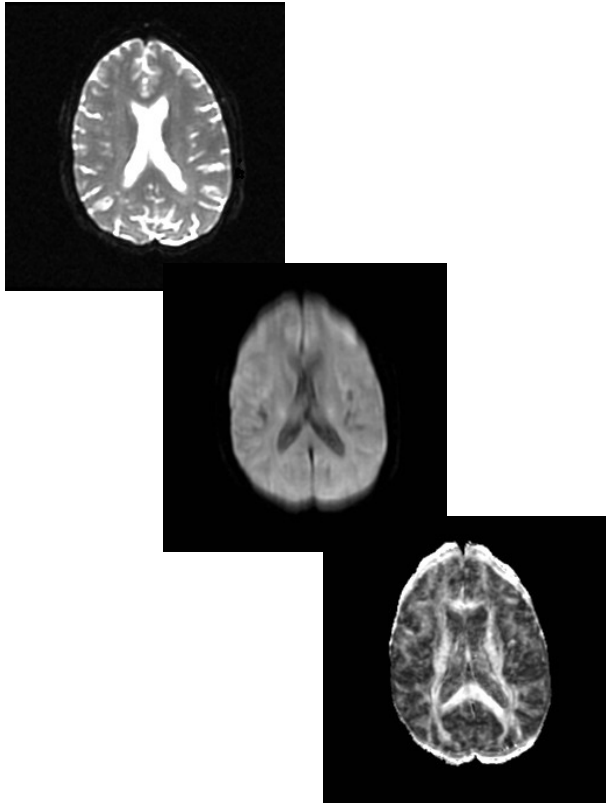
# Tensor Estimation

Adjust the **Window/Level** parameters of the baseline volume using the slider





Browse through the baseline images, which correspond to the volumes that have been acquired without gradient.



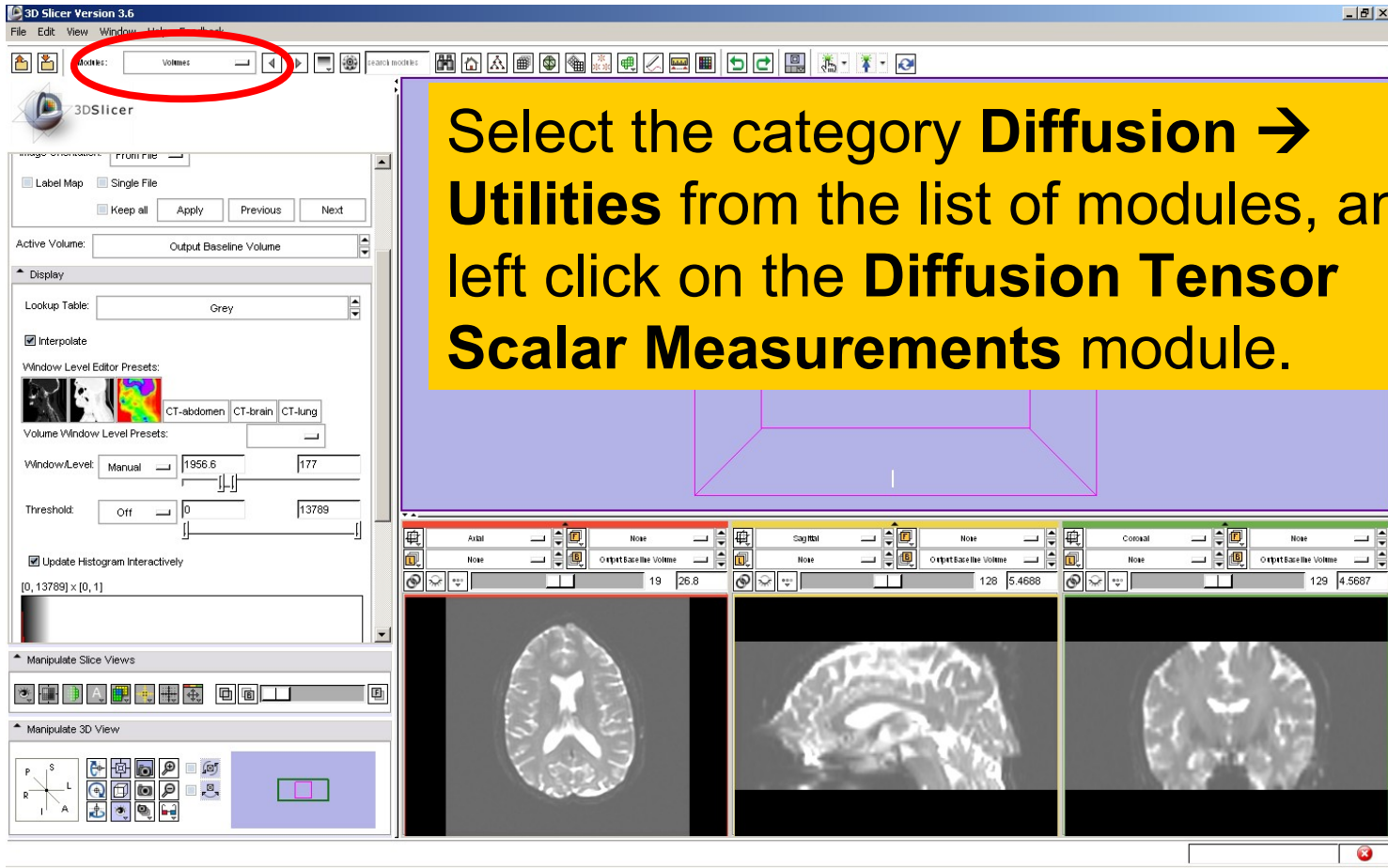
# Part 2:

# Scalar

# Measurements

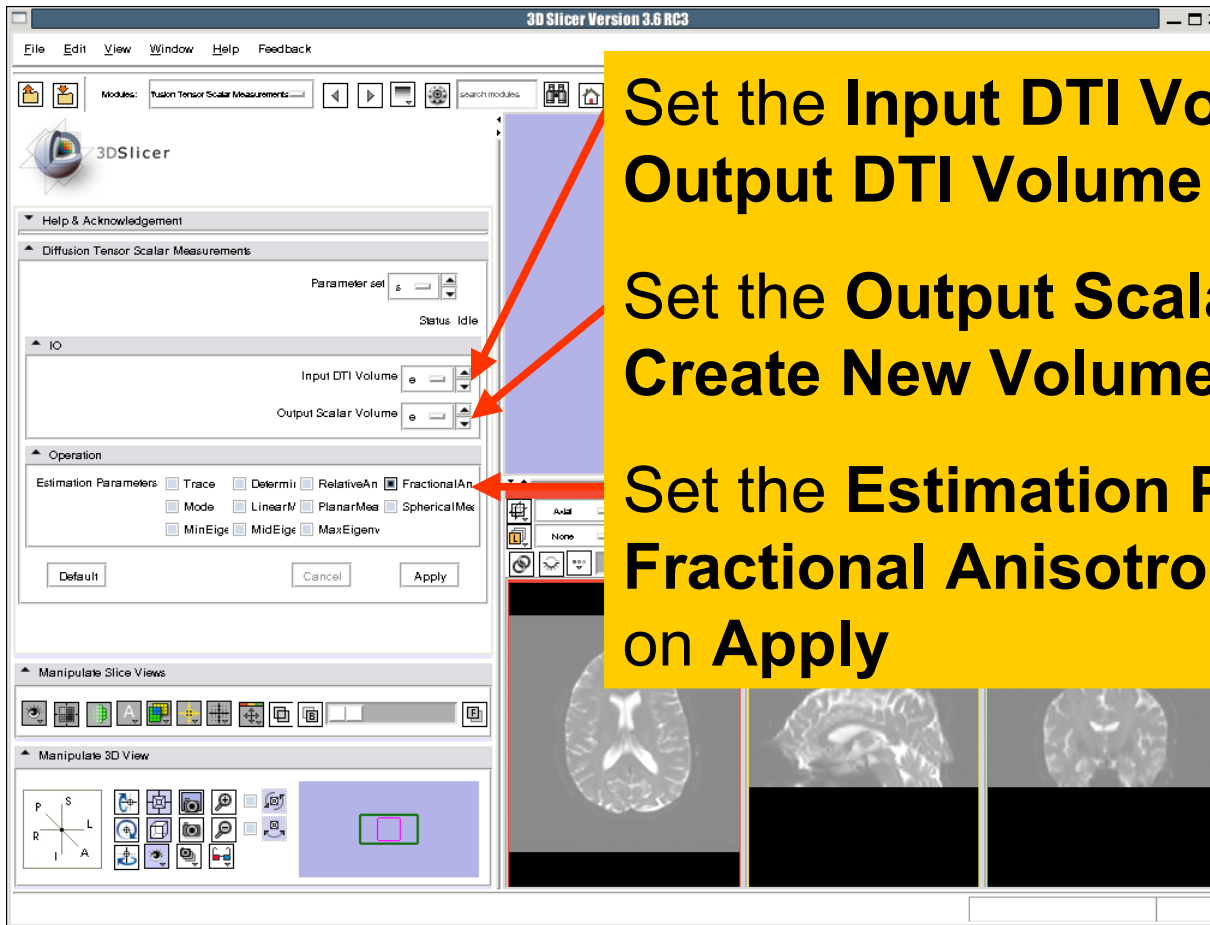


# Scalar Measurements





# Scalar Measurements



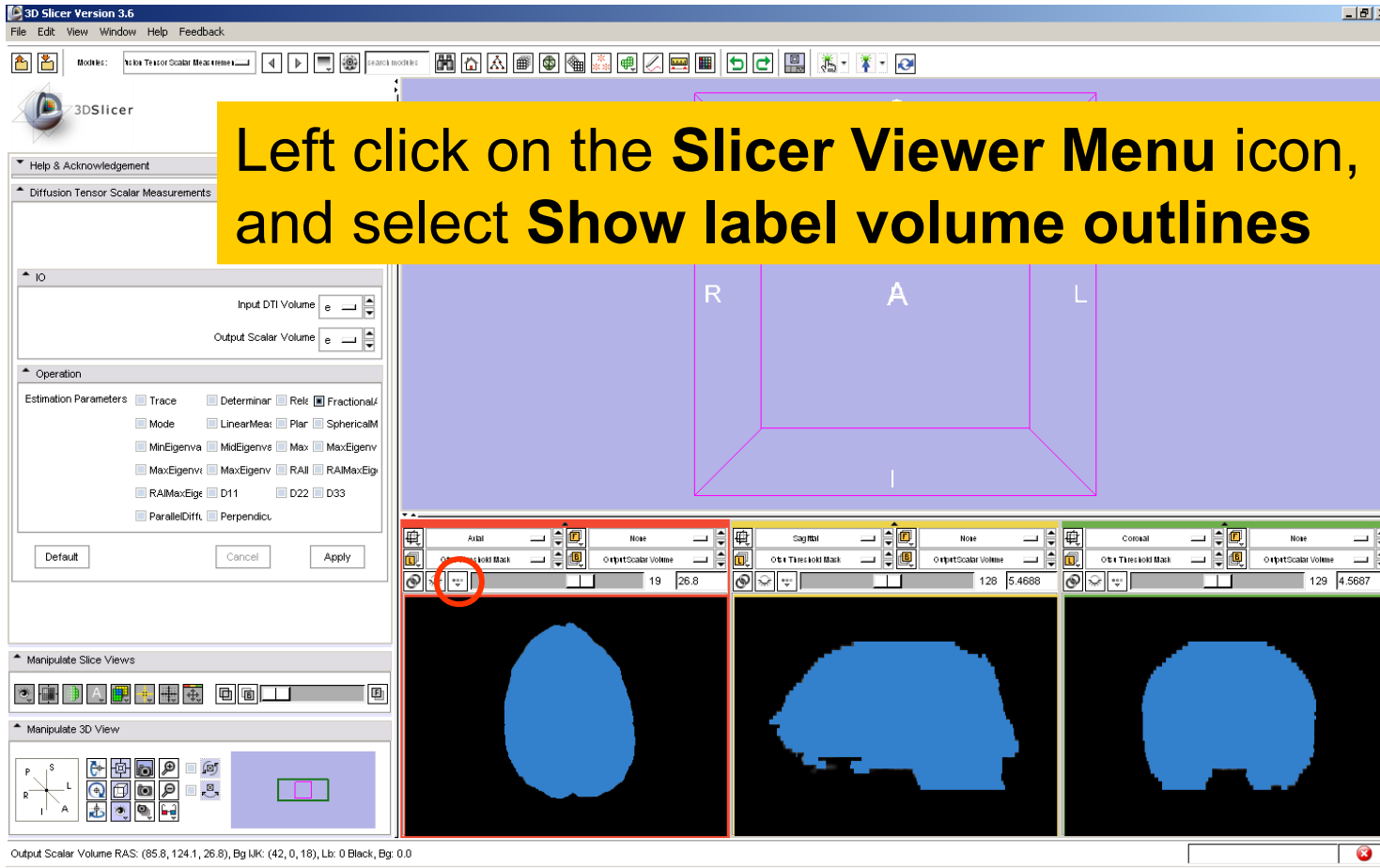
**Set the Input DTI Volume to  
Output DTI Volume**

**Set the Output Scalar Volume to  
Create New Volume**

**Set the Estimation Parameters to  
Fractional Anisotropy, and click  
on Apply**

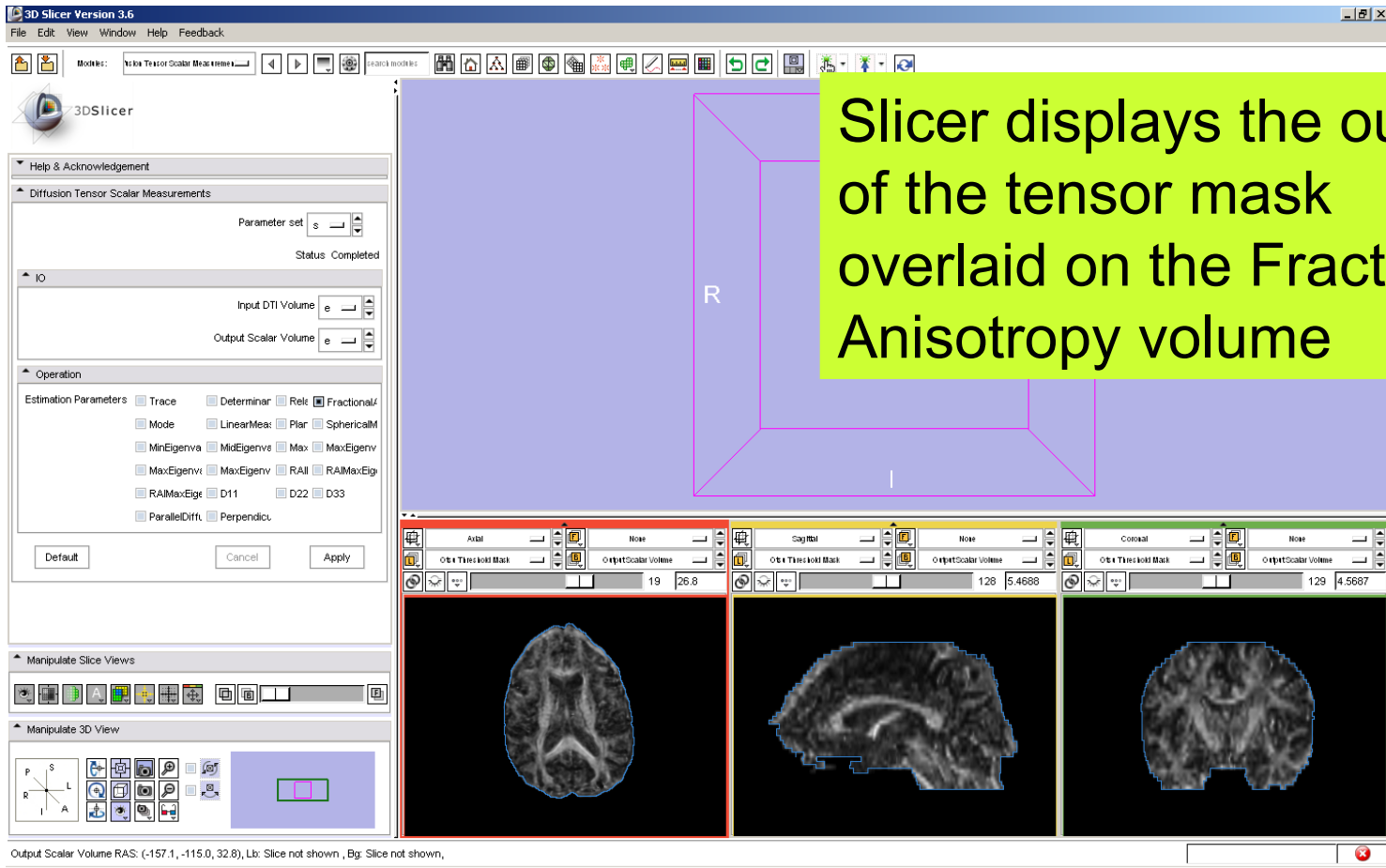


# Fractional Anisotropy Volume





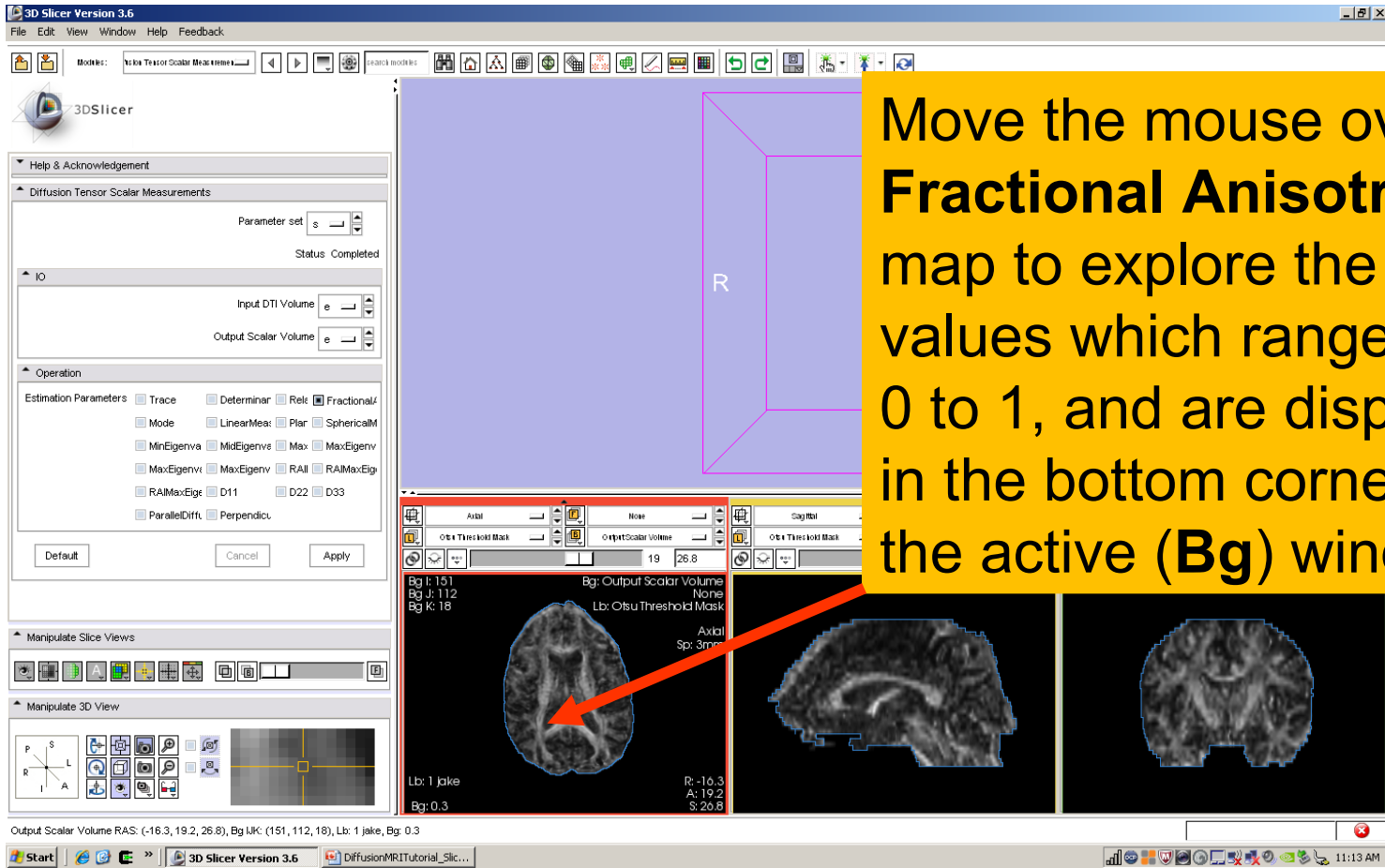
# Fractional Anisotropy Volume







# Fractional Anisotropy Volume



Move the mouse over the **Fractional Anisotropy** map to explore the FA values which range from 0 to 1, and are displayed in the bottom corner of the active (**Bg**) window.

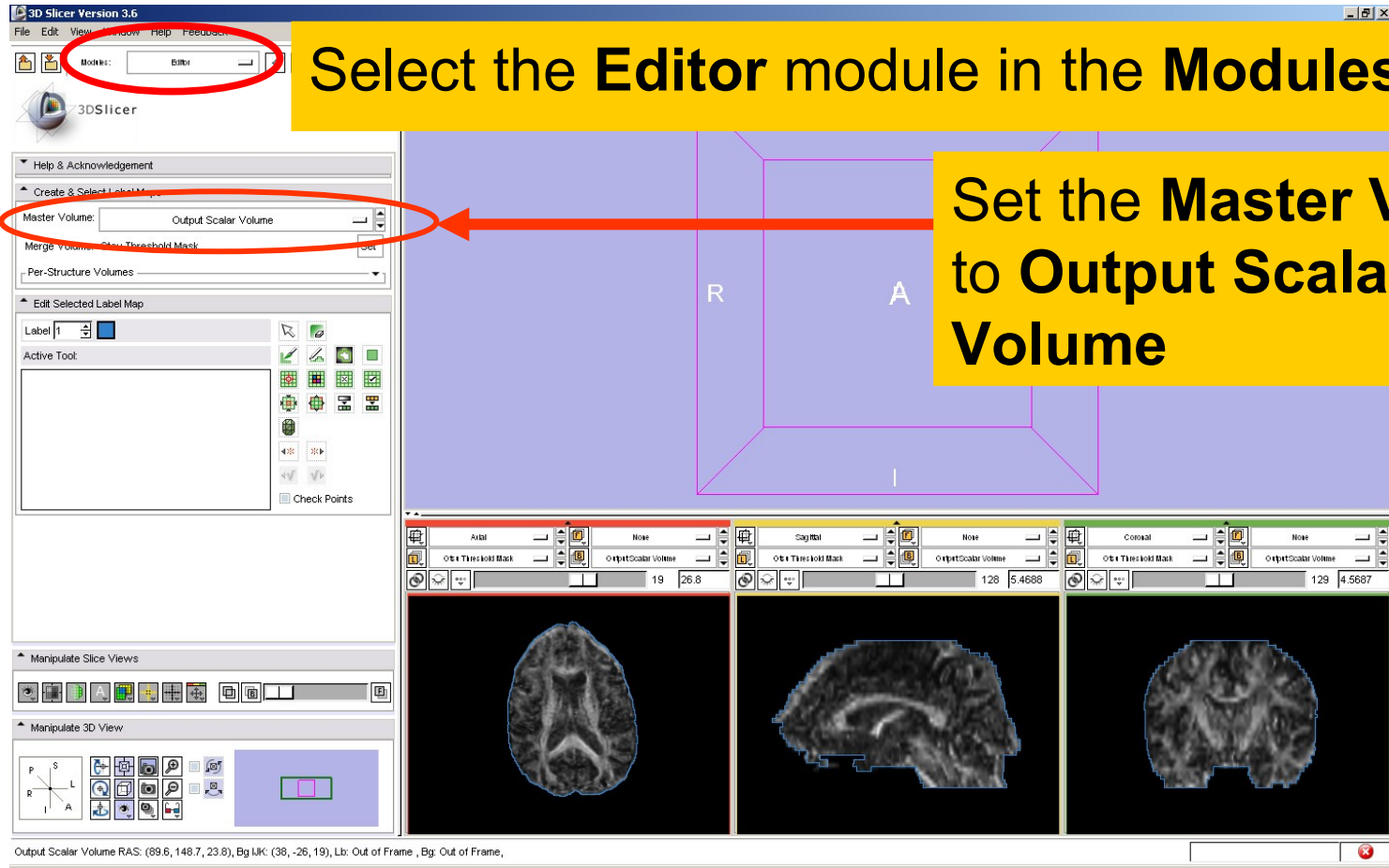


# **Part 3:**

## **Region of Interest Based Tractography**

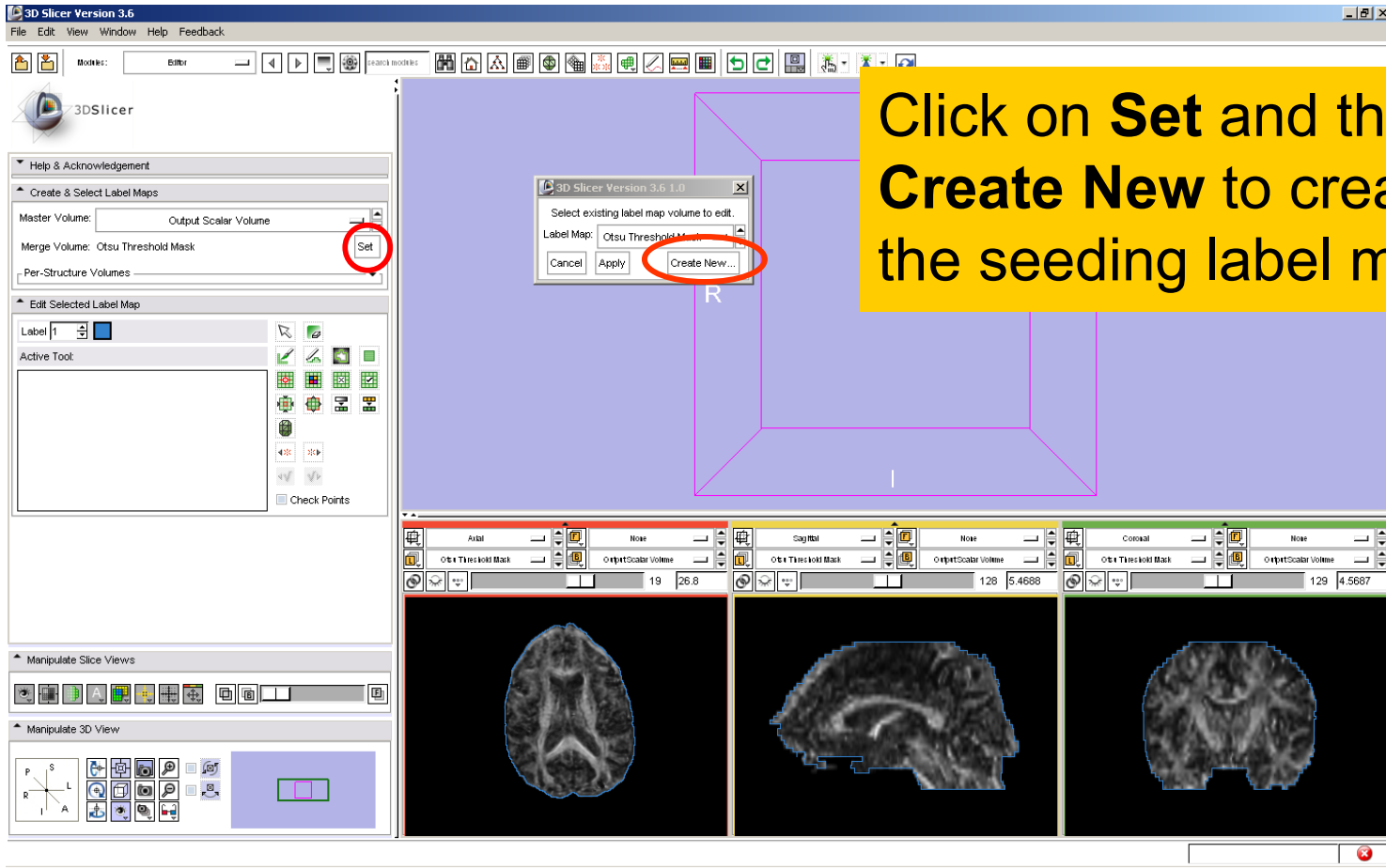


# LabelMap Generation



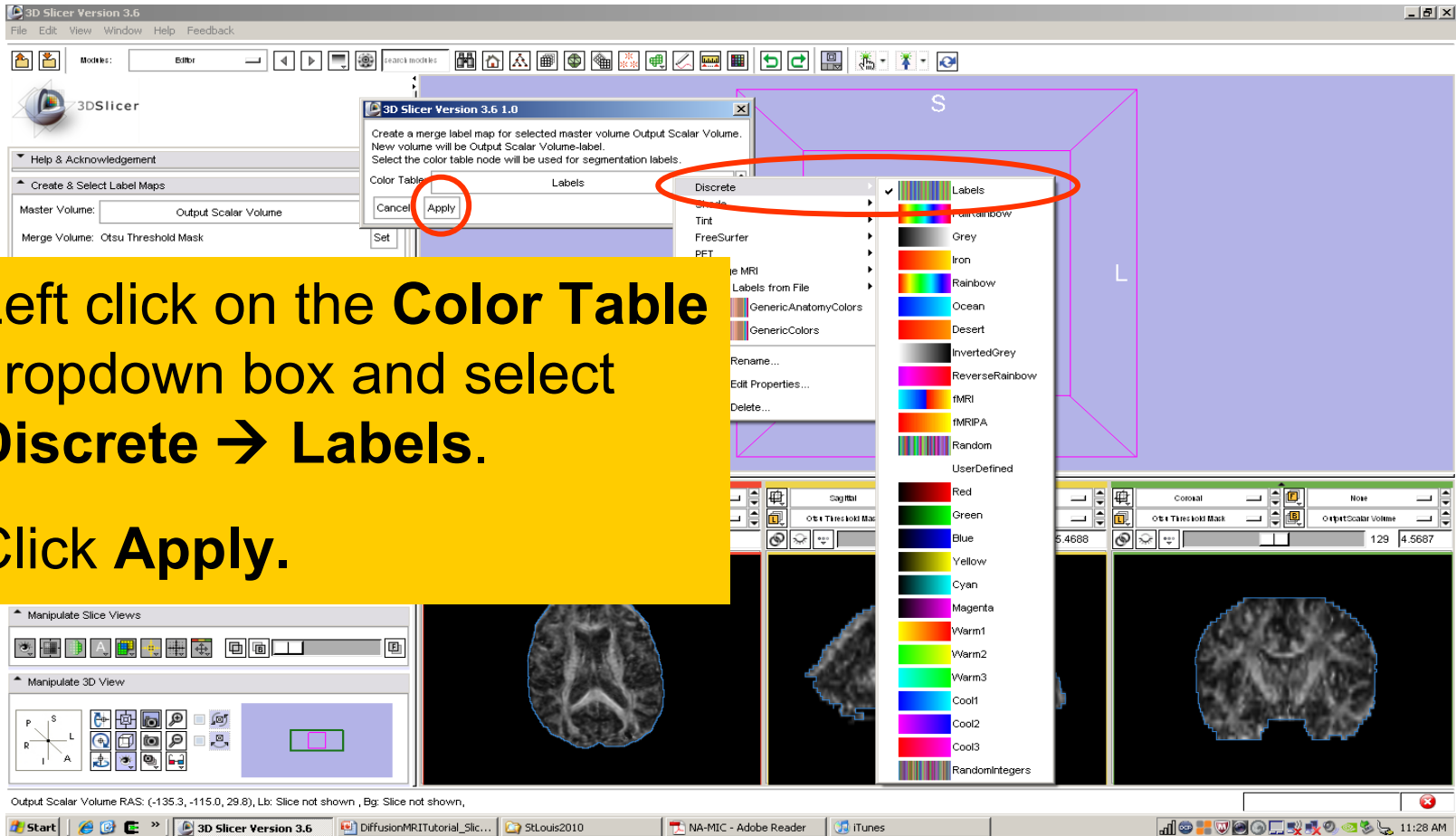


# LabelMap Generation



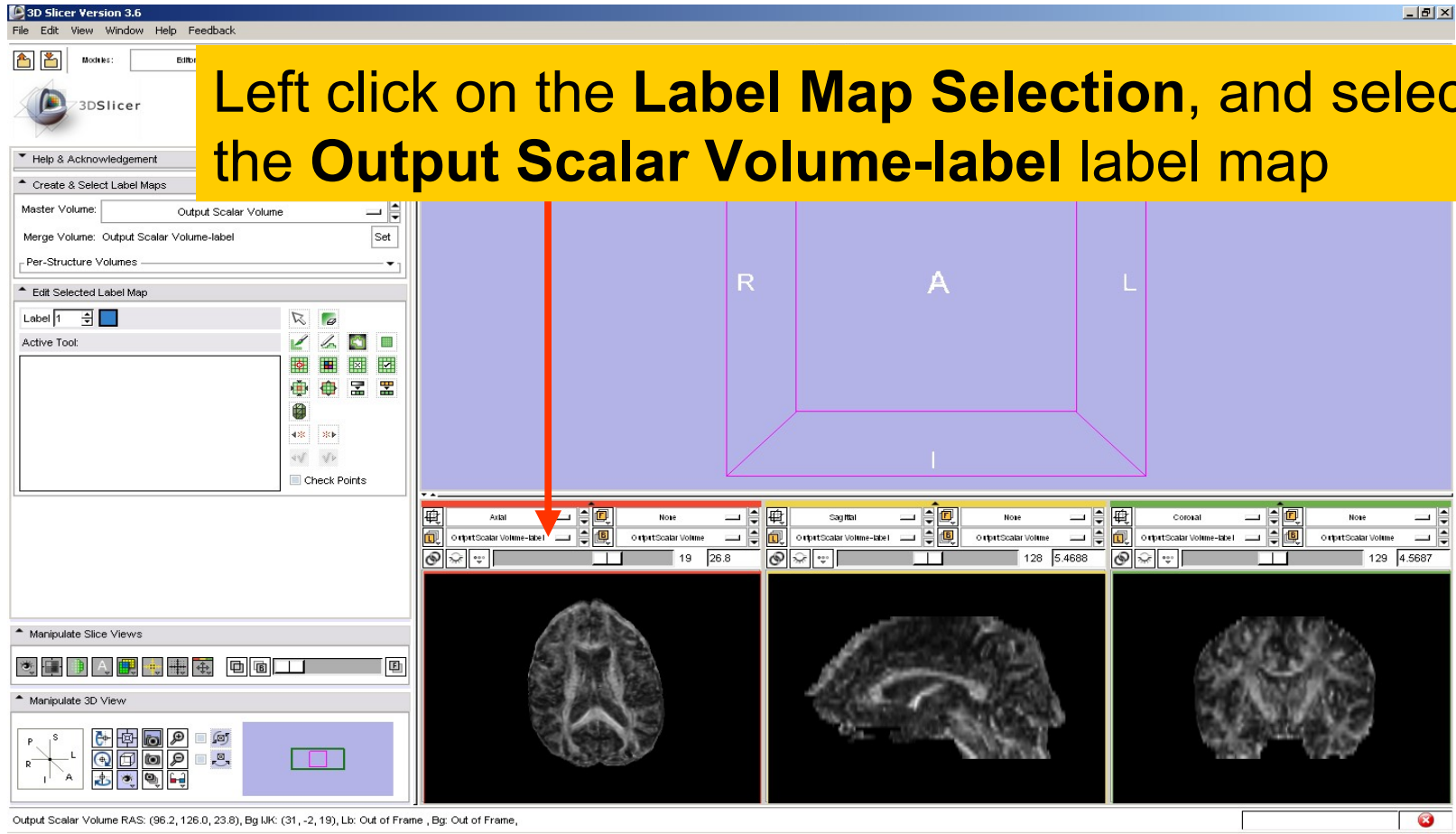


# LabelMap Generation



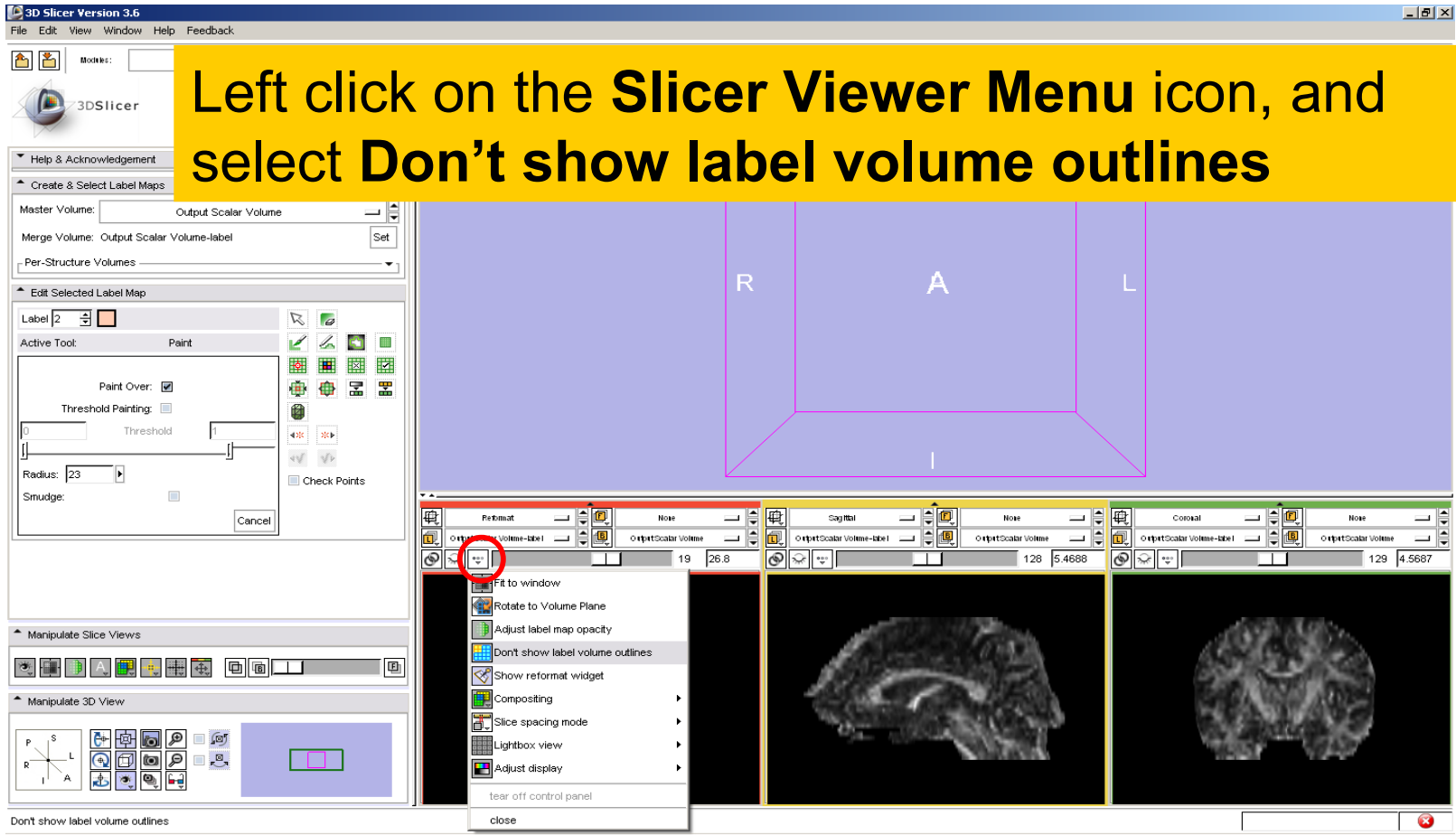


# LabelMap Generation





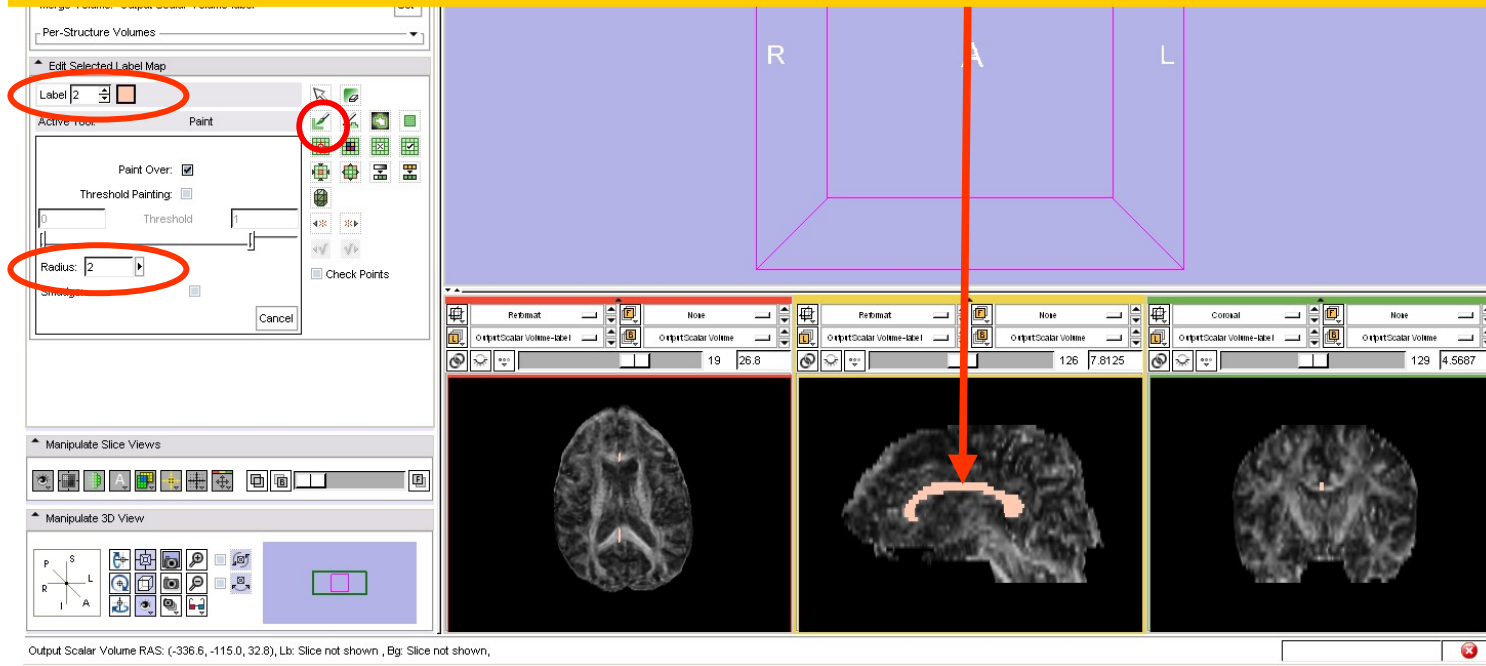
# LabelMap Generation





# LabelMap Generation

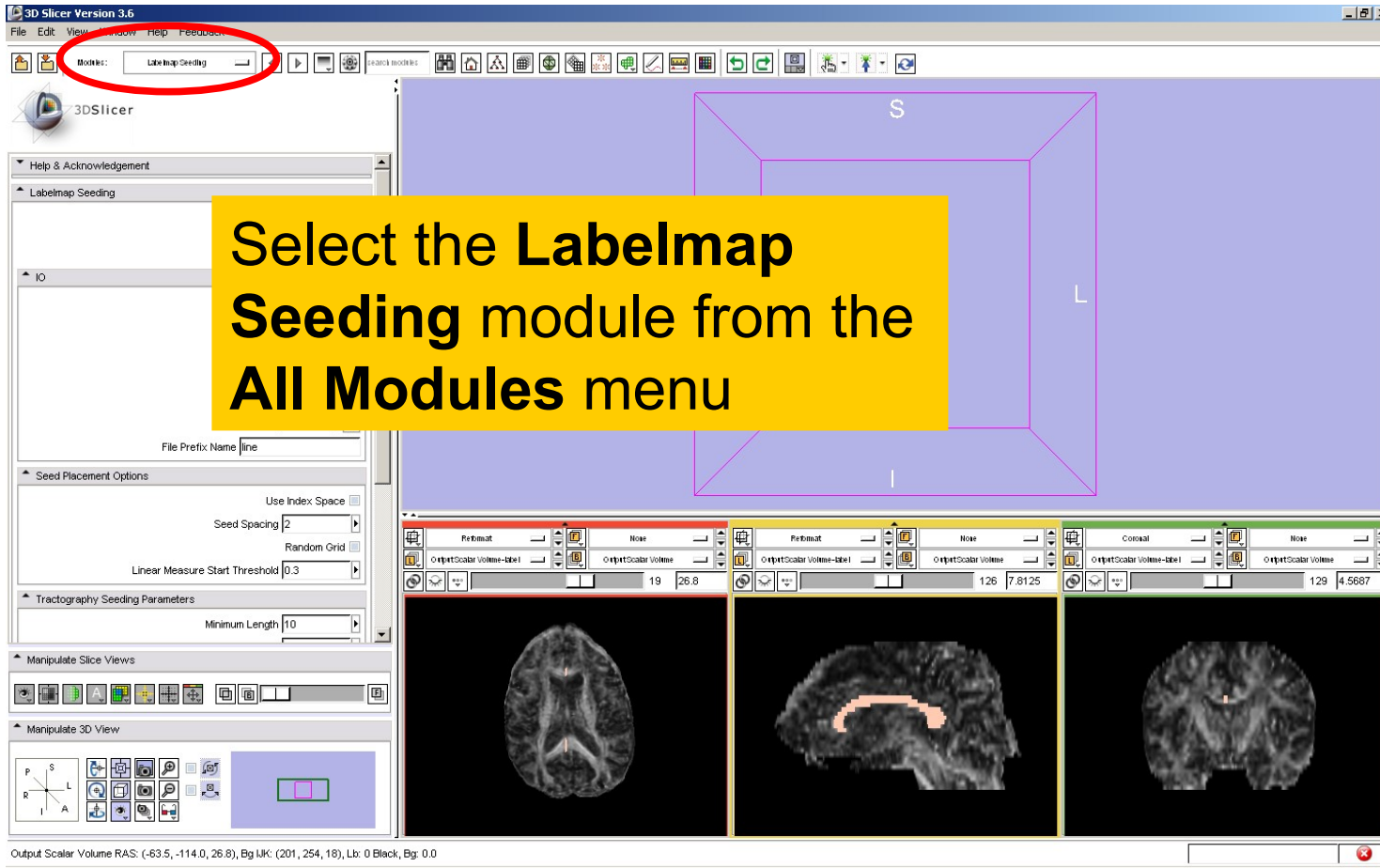
Select the label 2 (pink), click on the **Paint** icon, set the radius to **2** and draw a region of interest within the corpus callosum in the sagittal view on a set of 2 or 3 slices





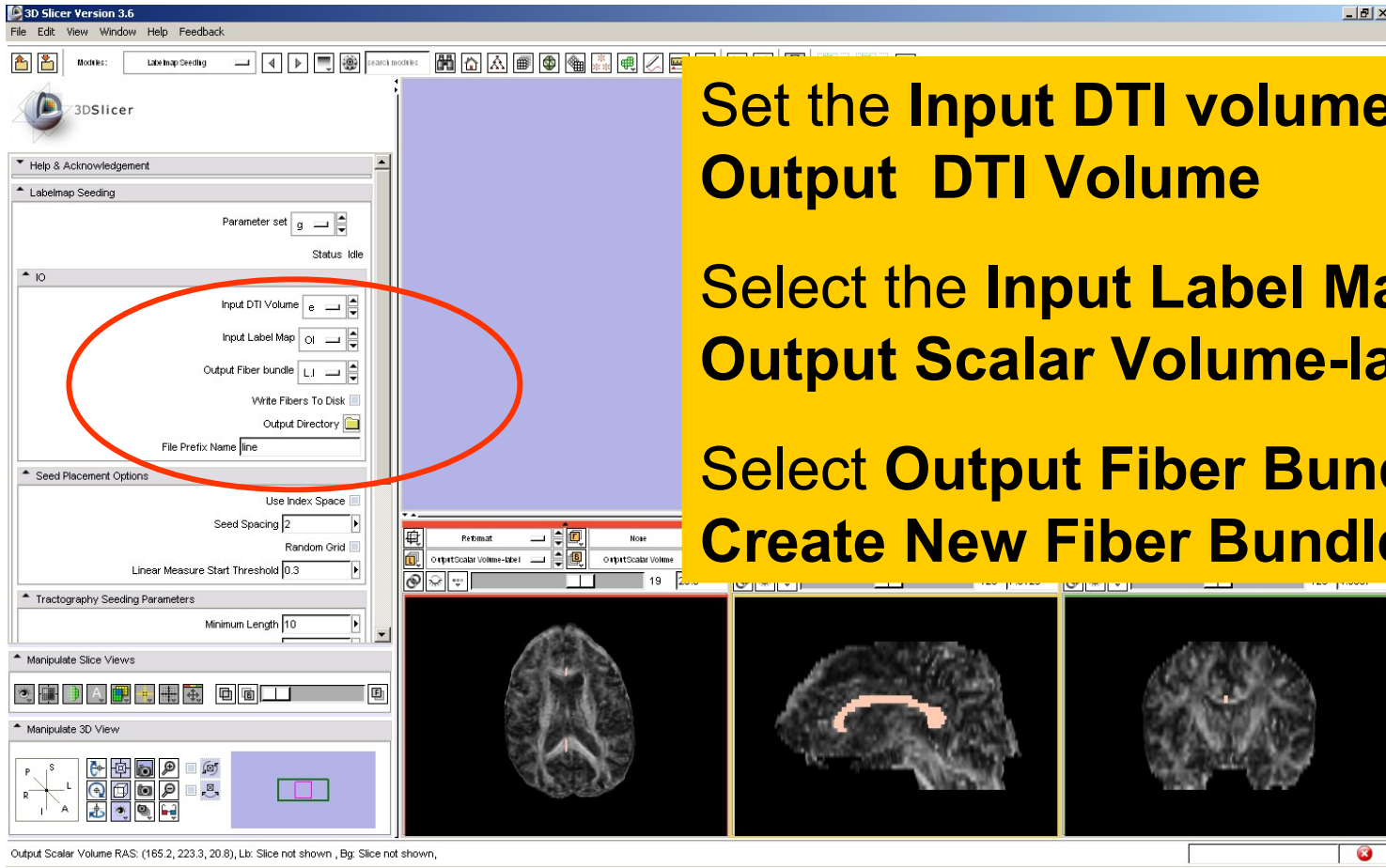


# LabelMap Seeding



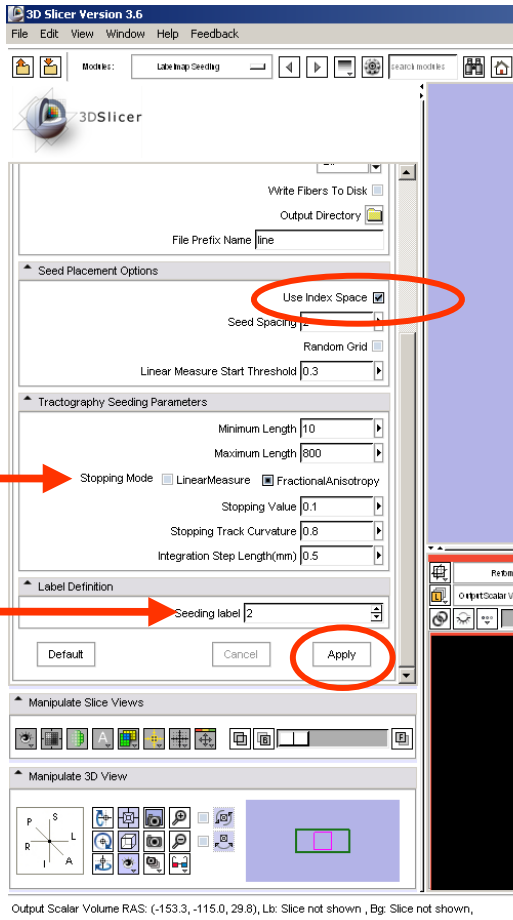


# LabelMap Seeding





# LabelMap Seeding



In the **Seed Placement Options** tab, check mark **Use Index Space**.

In the **Tractography Seeding Parameters** tab, set the **Stopping Mode** to **Fractional Anisotropy**, and use the default parameters for the Minimum and Maximum Length, Stopping Value, Stopping Track Curvature and Integration Step Length.

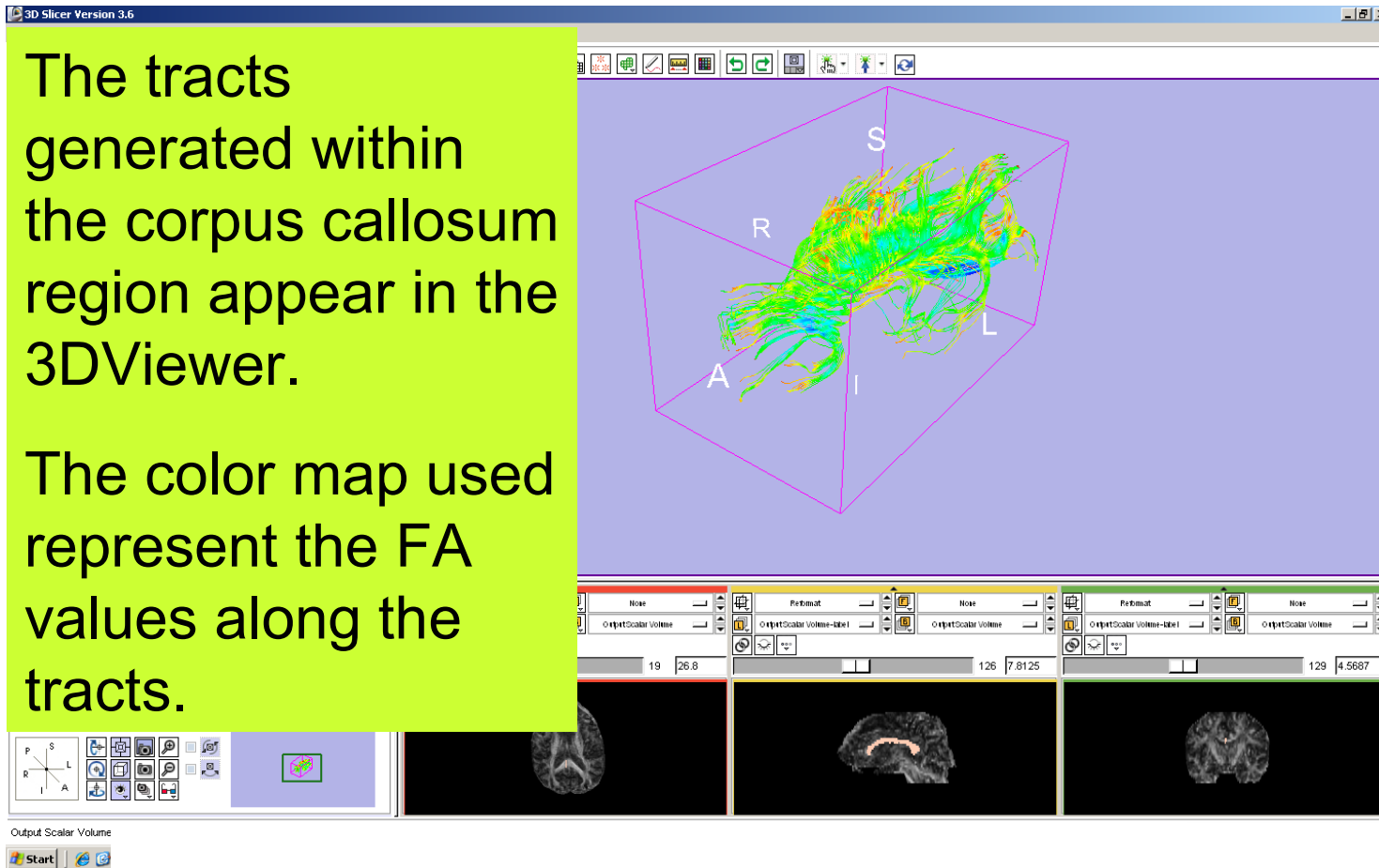
In the **Label Definition** tab, set Seeding label to label 2, and click on **Apply**.



# LabelMap Seeding

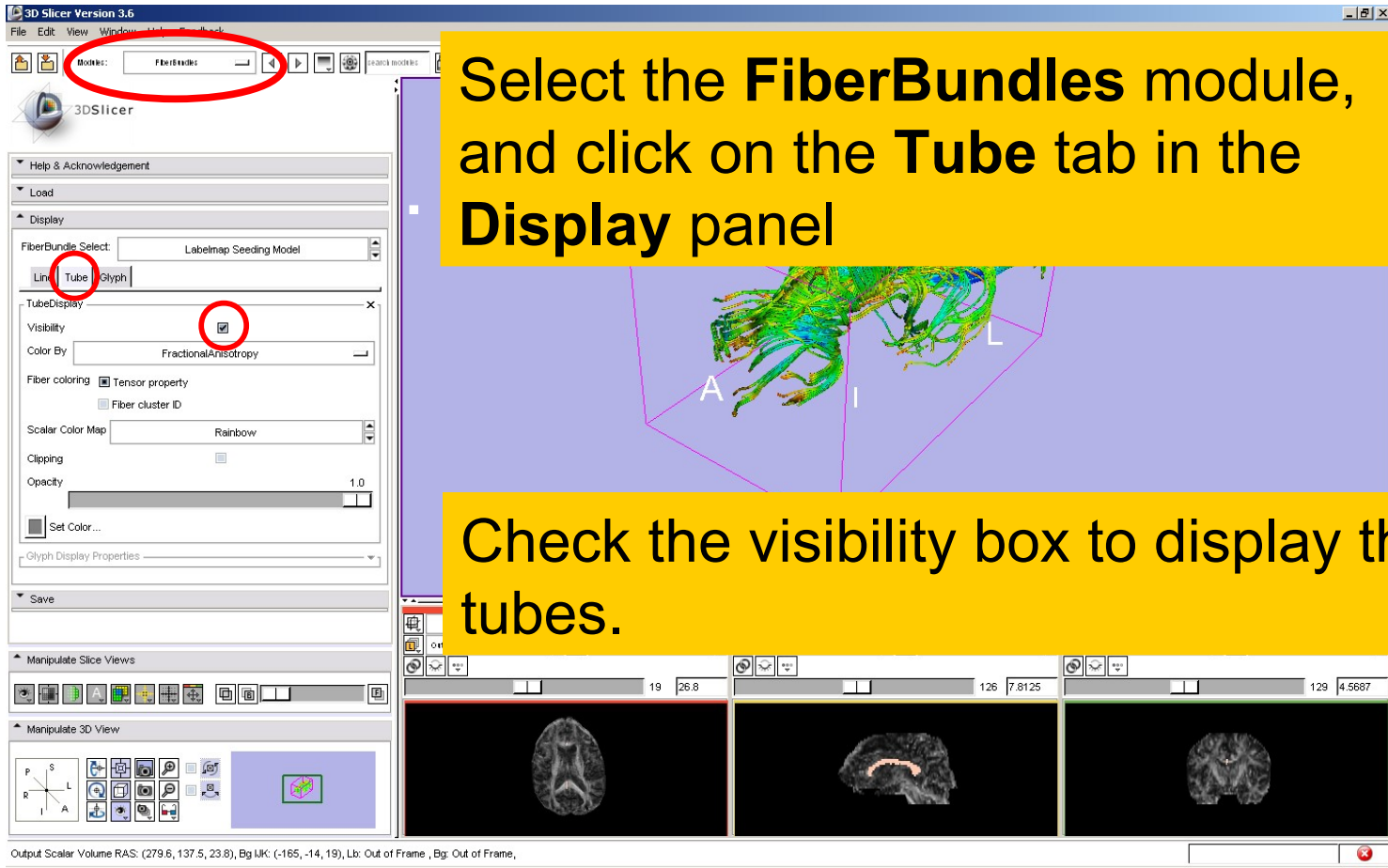
The tracts generated within the corpus callosum region appear in the 3DViewer.

The color map used represent the FA values along the tracts.





# LabelMap Seeding



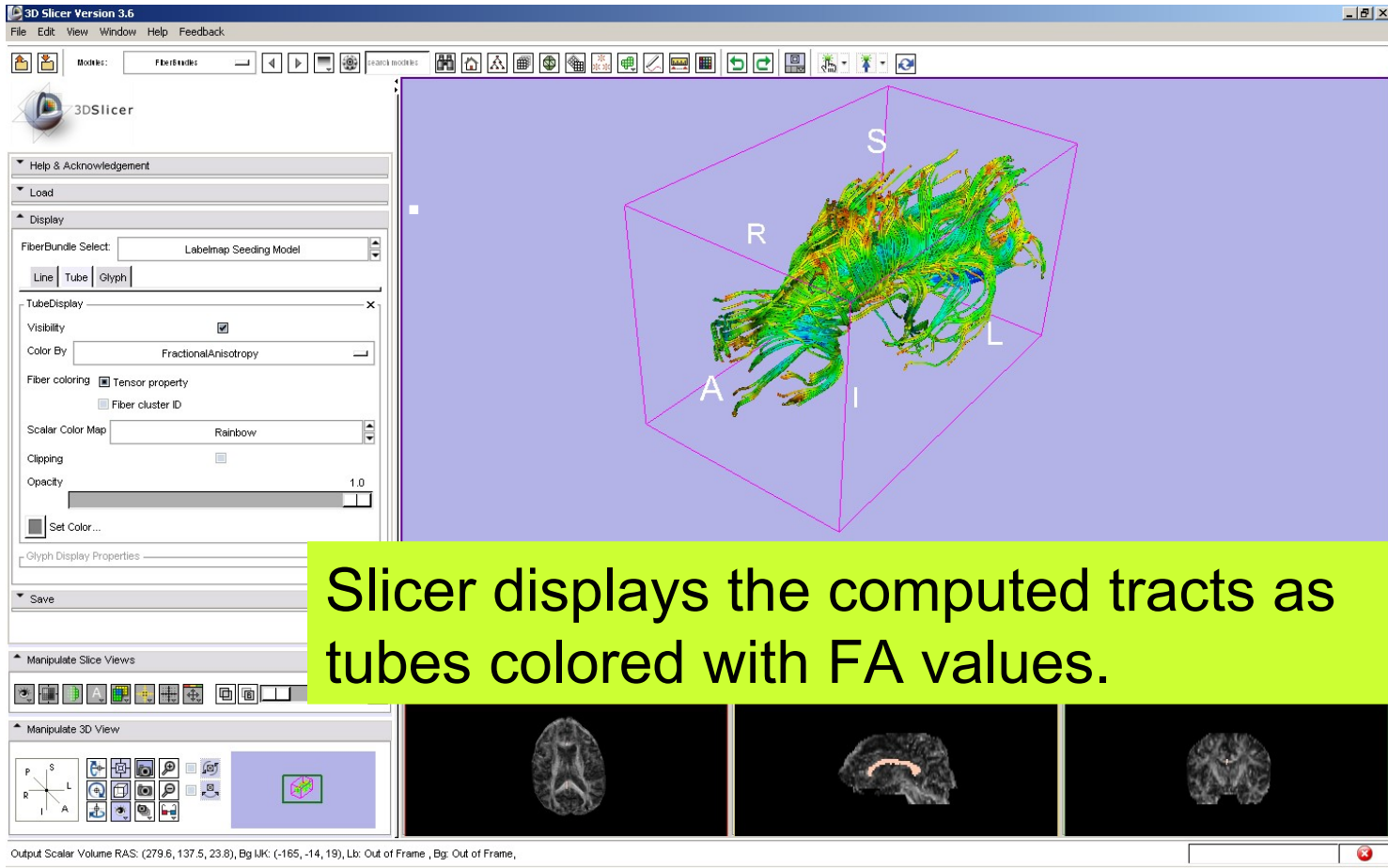
Select the **FiberBundles** module, and click on the **Tube** tab in the **Display** panel

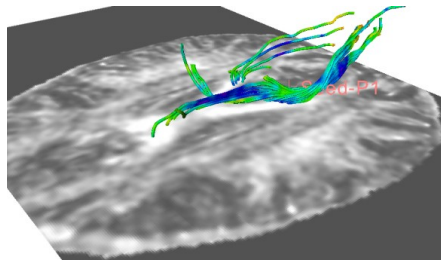
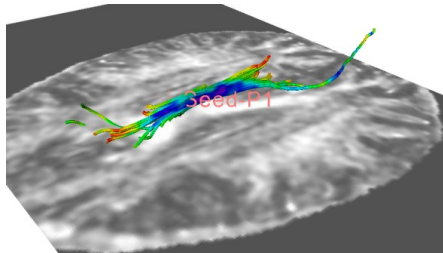
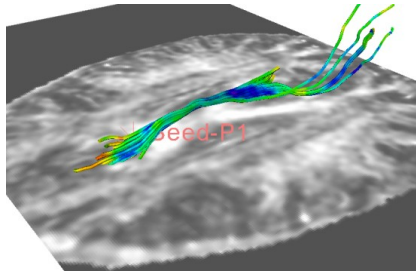
Check the visibility box to display the tubes.

Output Scalar Volume RAS: (279.6, 137.5, 23.8), Bg IJK: (-165, -14, 19), Lb: Out of Frame, Bg: Out of Frame,



# LabelMap Seeding





## Part 4:

# Tractography on-the-fly

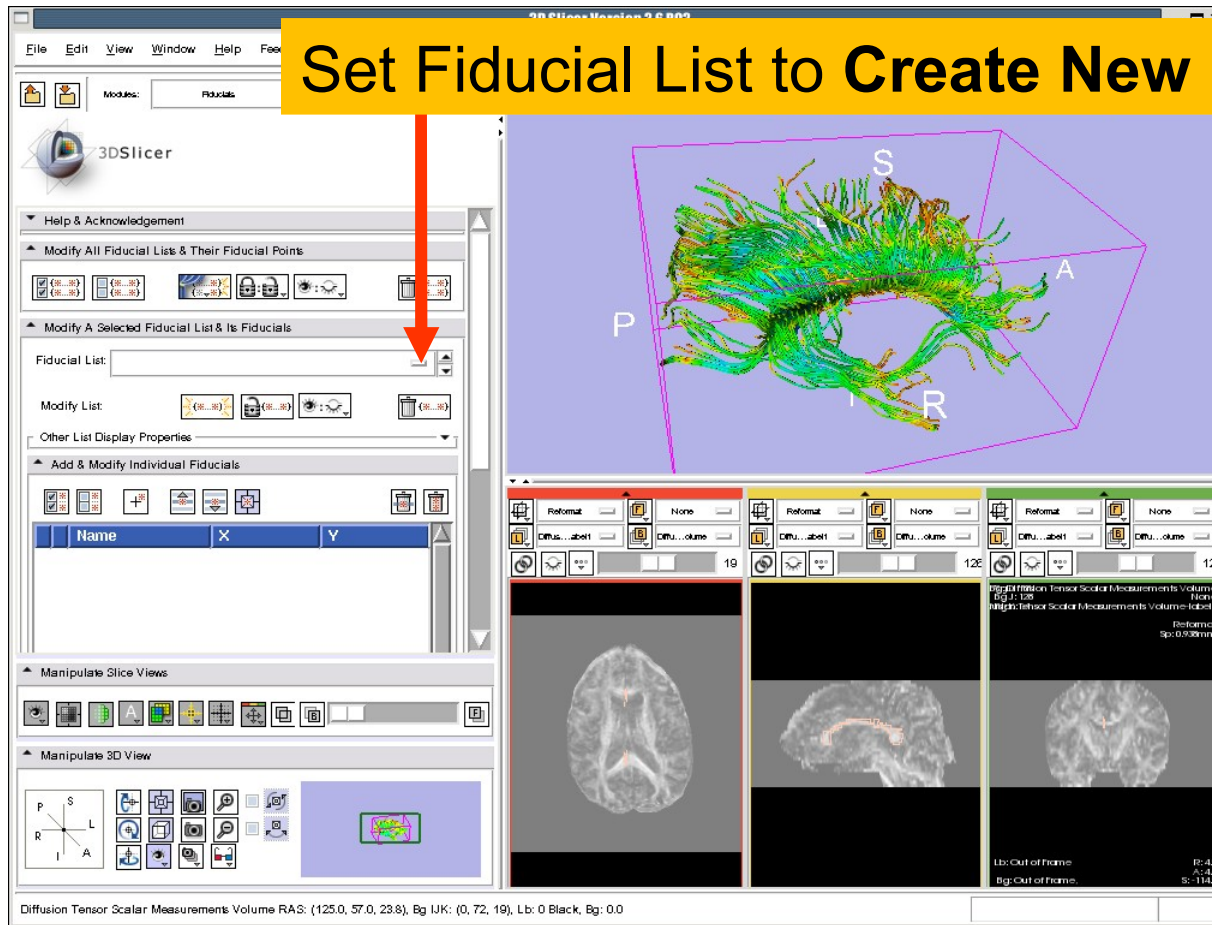






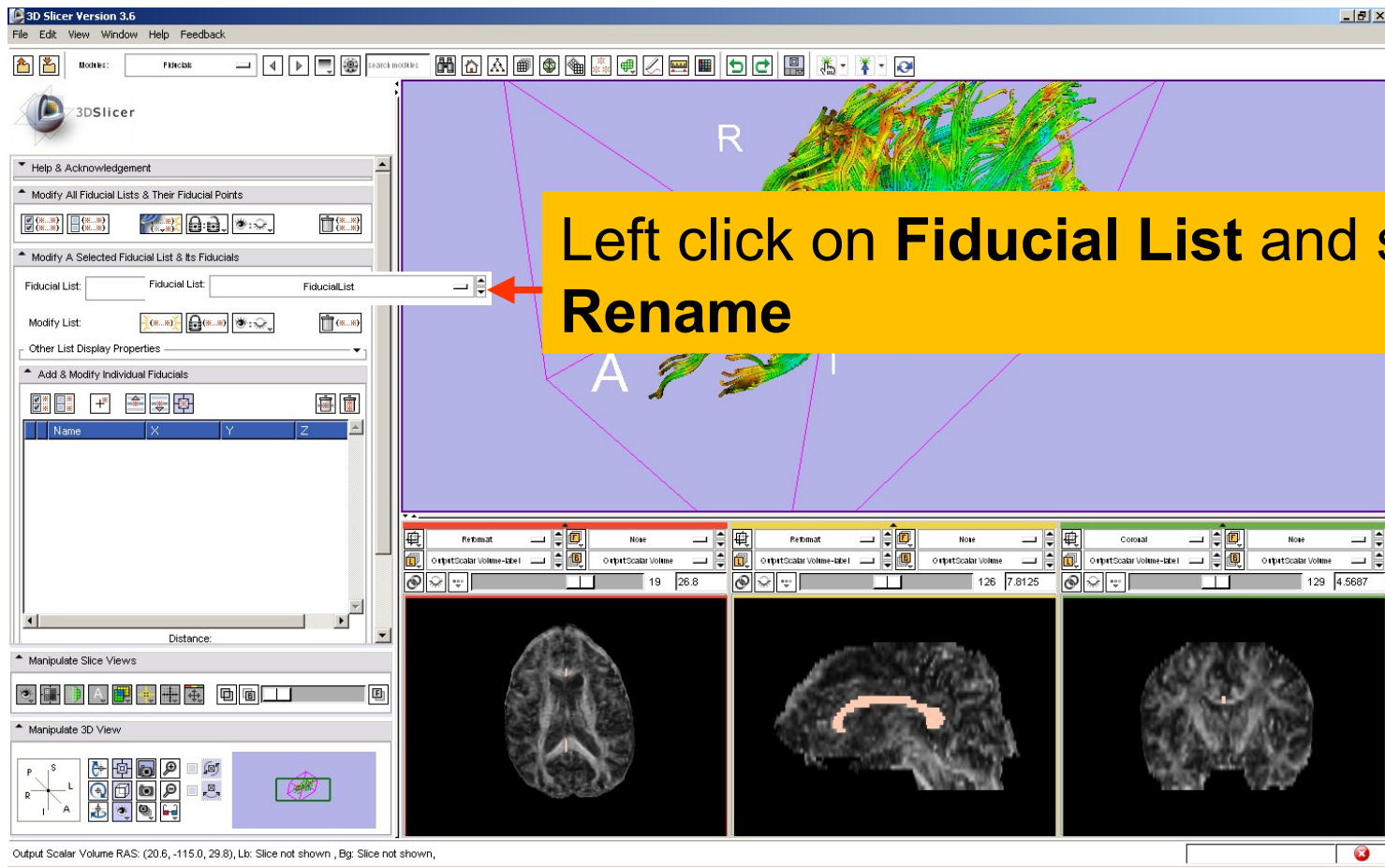


# Fiducial Seeding





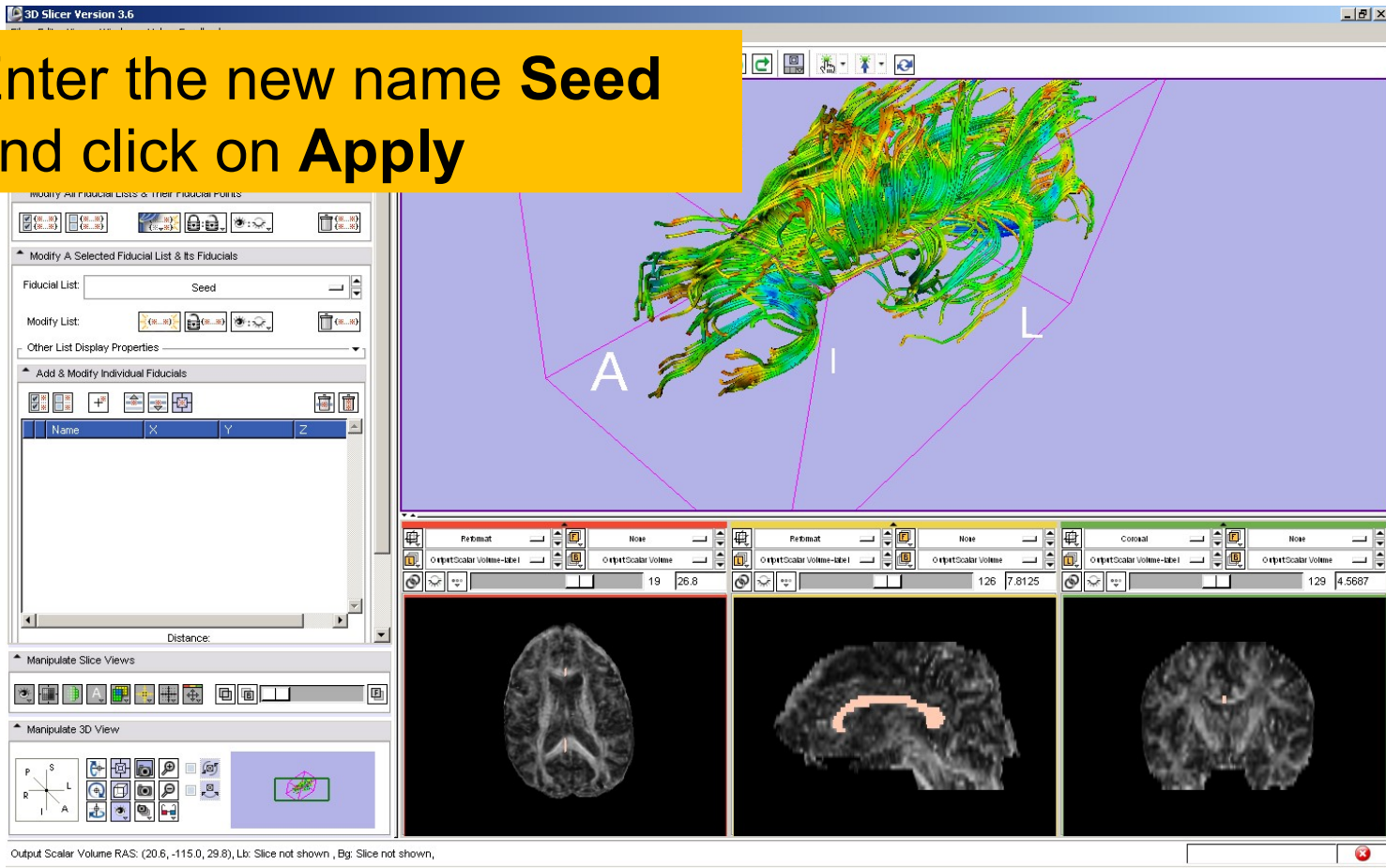
# Fiducial Seeding





# Fiducial Seeding

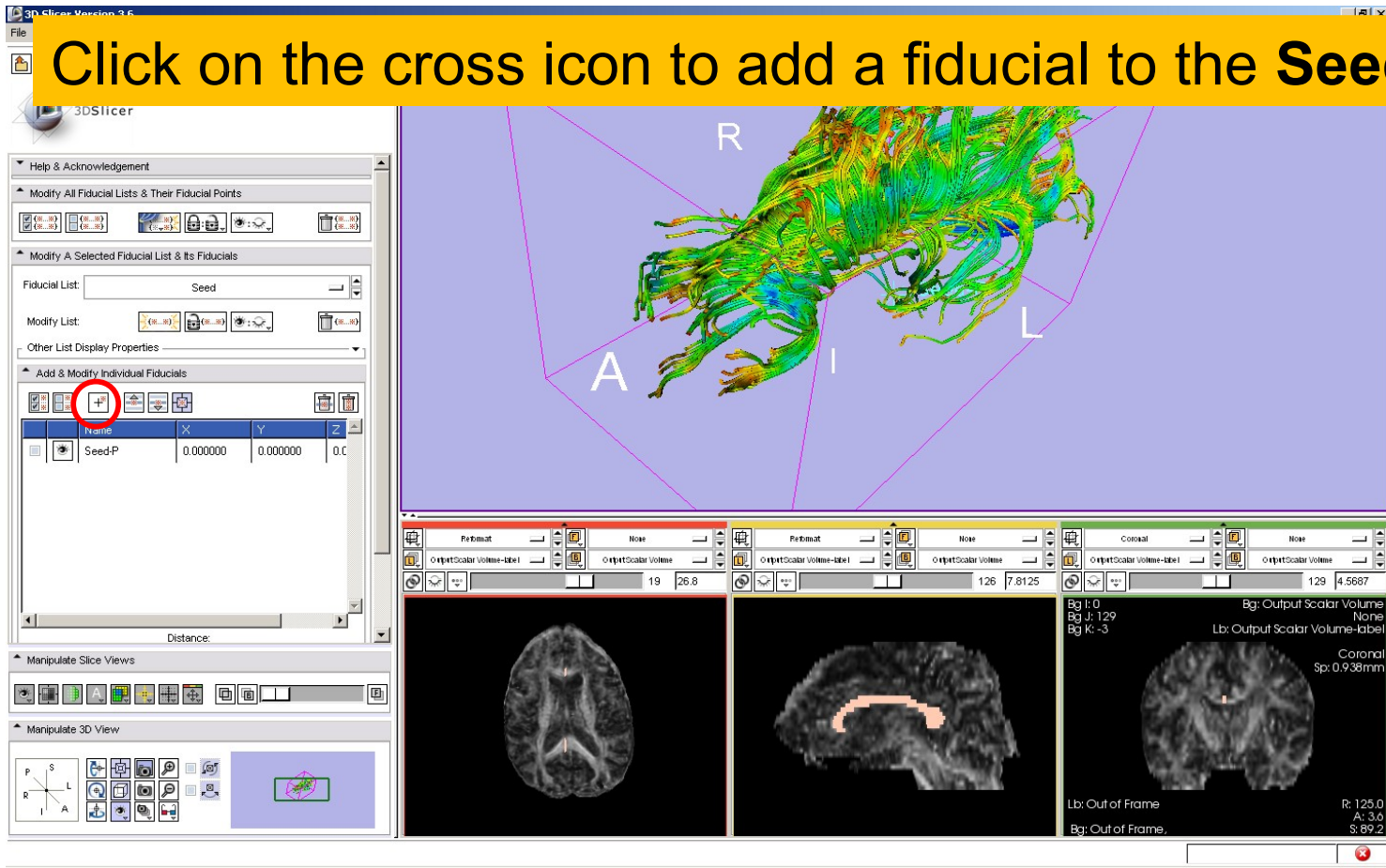
Enter the new name **Seed**  
and click on **Apply**





# Fiducial Seeding

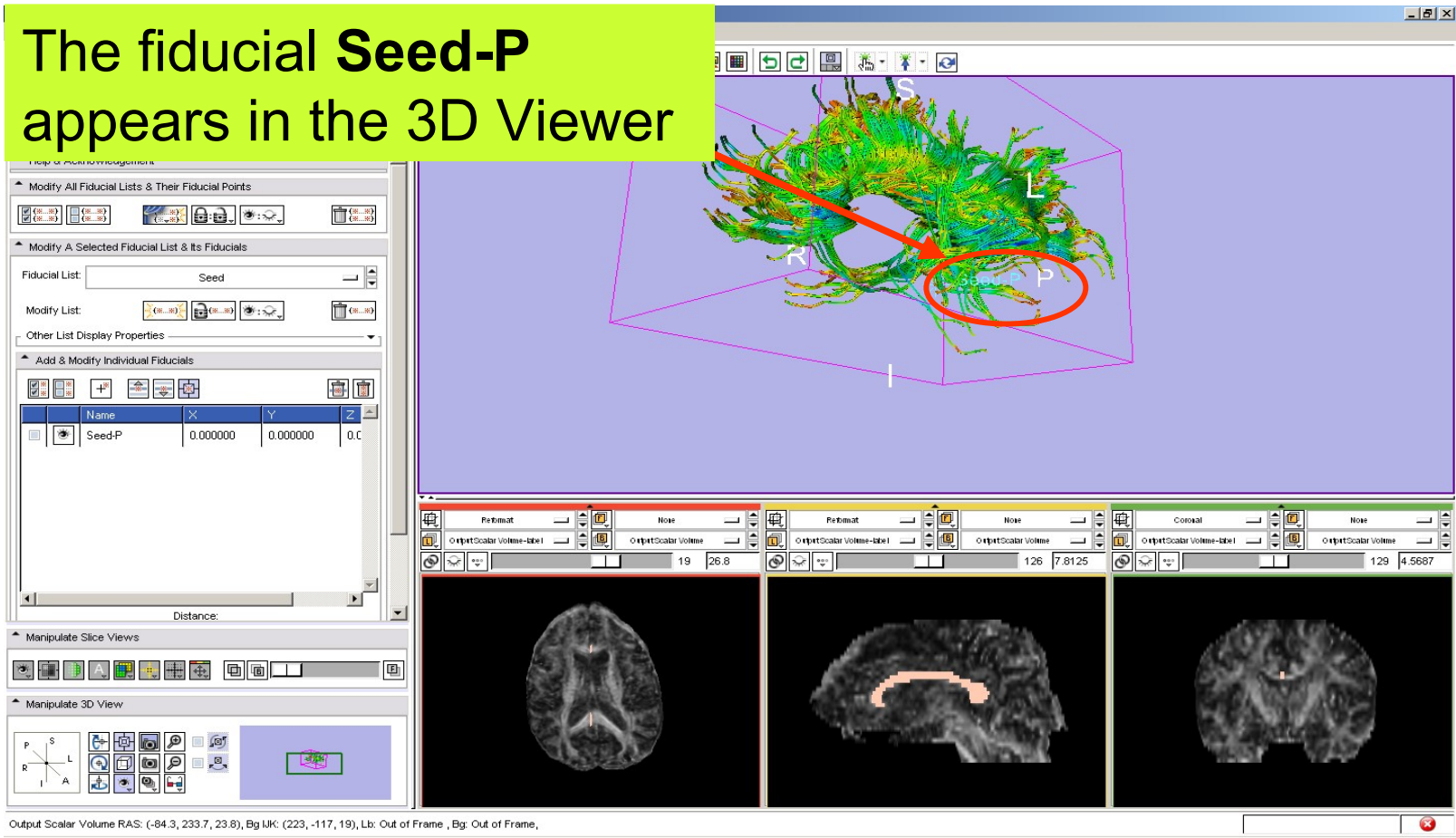
Click on the cross icon to add a fiducial to the **Seed** list





# Fiducial Seeding

The fiducial **Seed-P** appears in the 3D Viewer

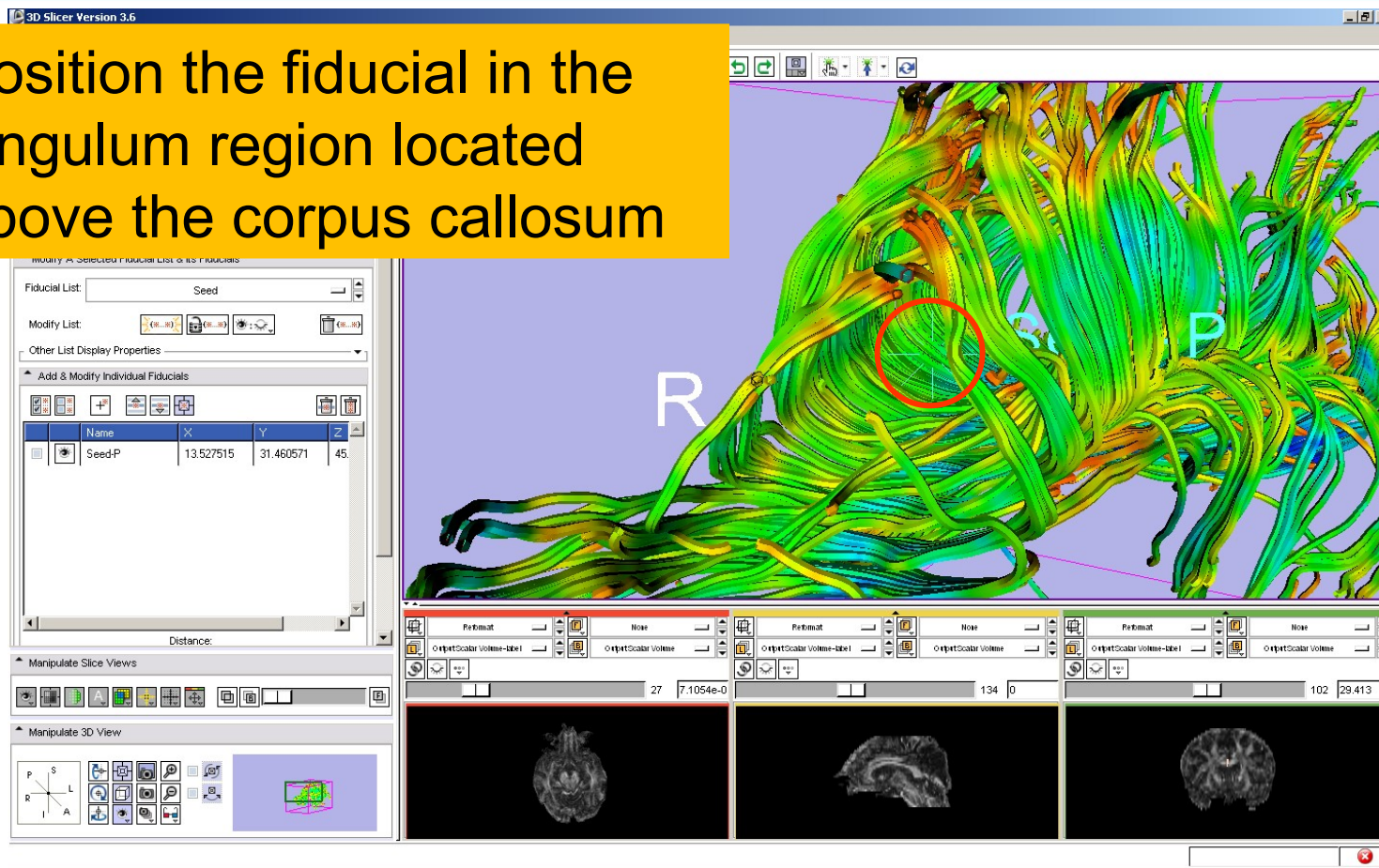






# Fiducial Seeding

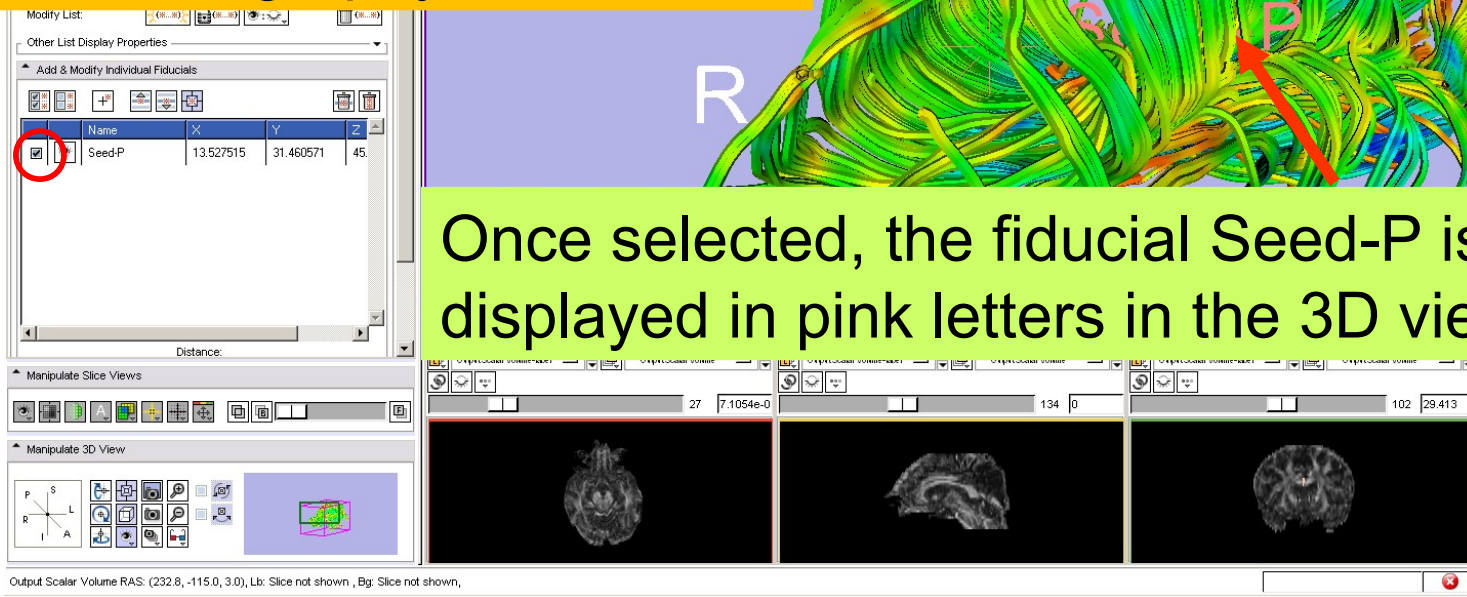
Position the fiducial in the cingulum region located above the corpus callosum





# Fiducial Seeding

**Check the box to select the 'Seed-P' fiducial:** we will use this fiducial to drive the tractography





# Fiducial Seeding

**Set the Output FiberBundleNode to Create New FiberBundle**

**Important:** this step **must** be done first

Select the **Fiducial Seeding** module

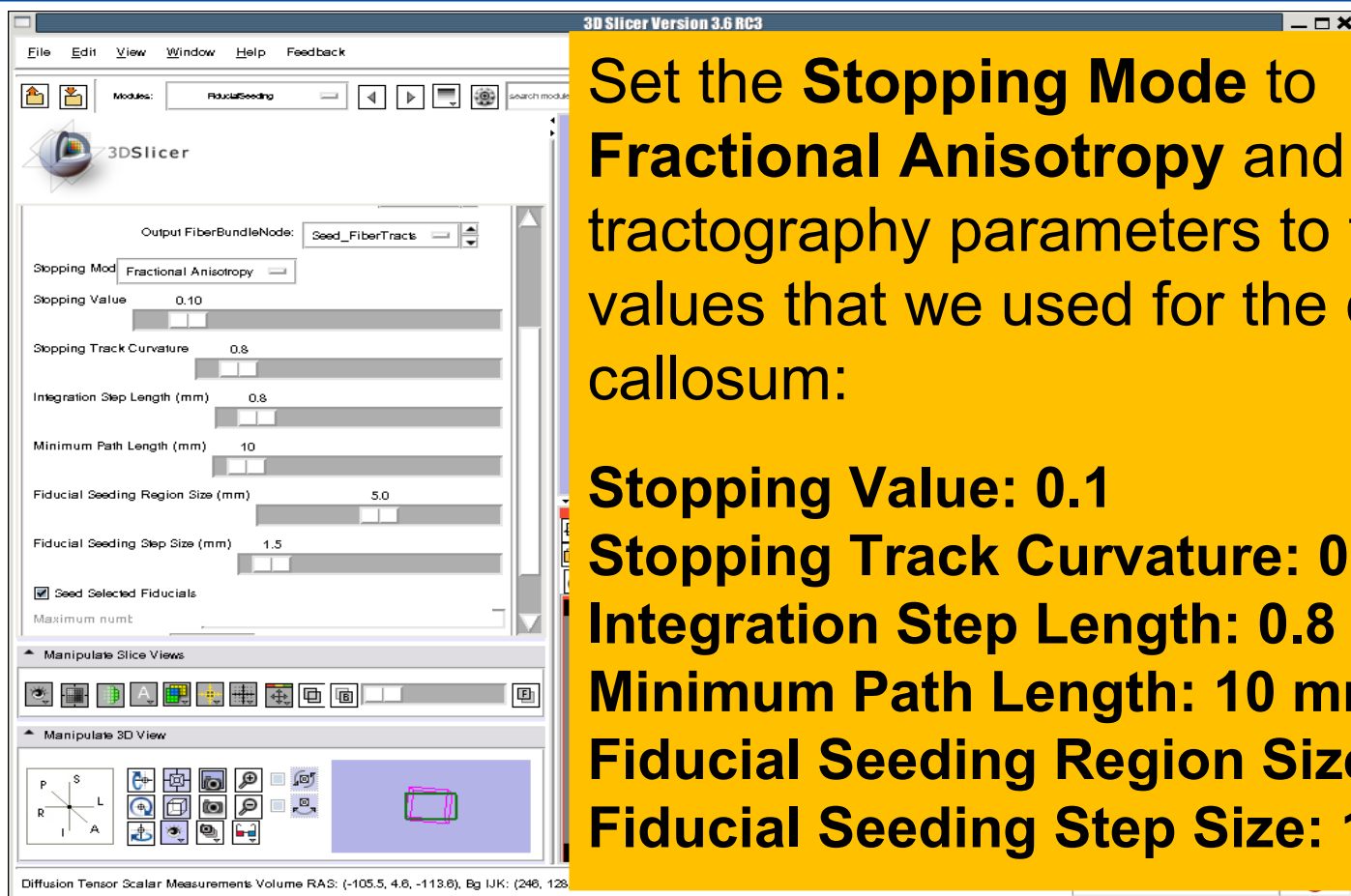
Set the DTI Volume to **Output DTI Volume**

Select Fiducial List: **Seed**





# Fiducial Seeding



**Set the Stopping Mode to Fractional Anisotropy and set the tractography parameters to the values that we used for the corpus callosum:**

**Stopping Value: 0.1**

**Stopping Track Curvature: 0.8**

**Integration Step Length: 0.8 mm**

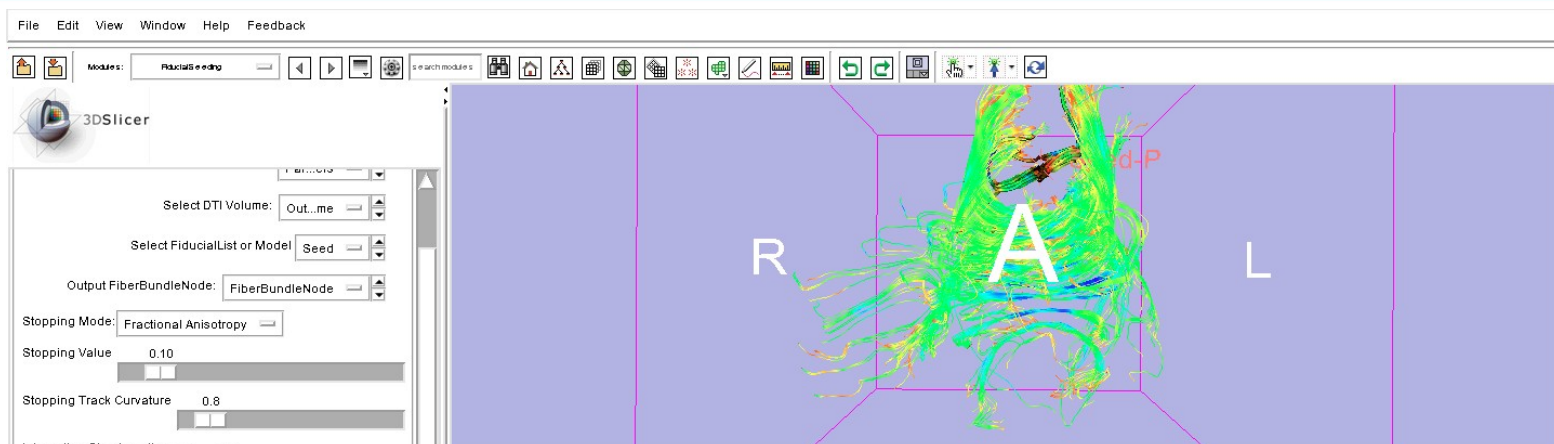
**Minimum Path Length: 10 mm**

**Fiducial Seeding Region Size: 5 mm**

**Fiducial Seeding Step Size: 1.5 mm**



# Fiducial Seeding



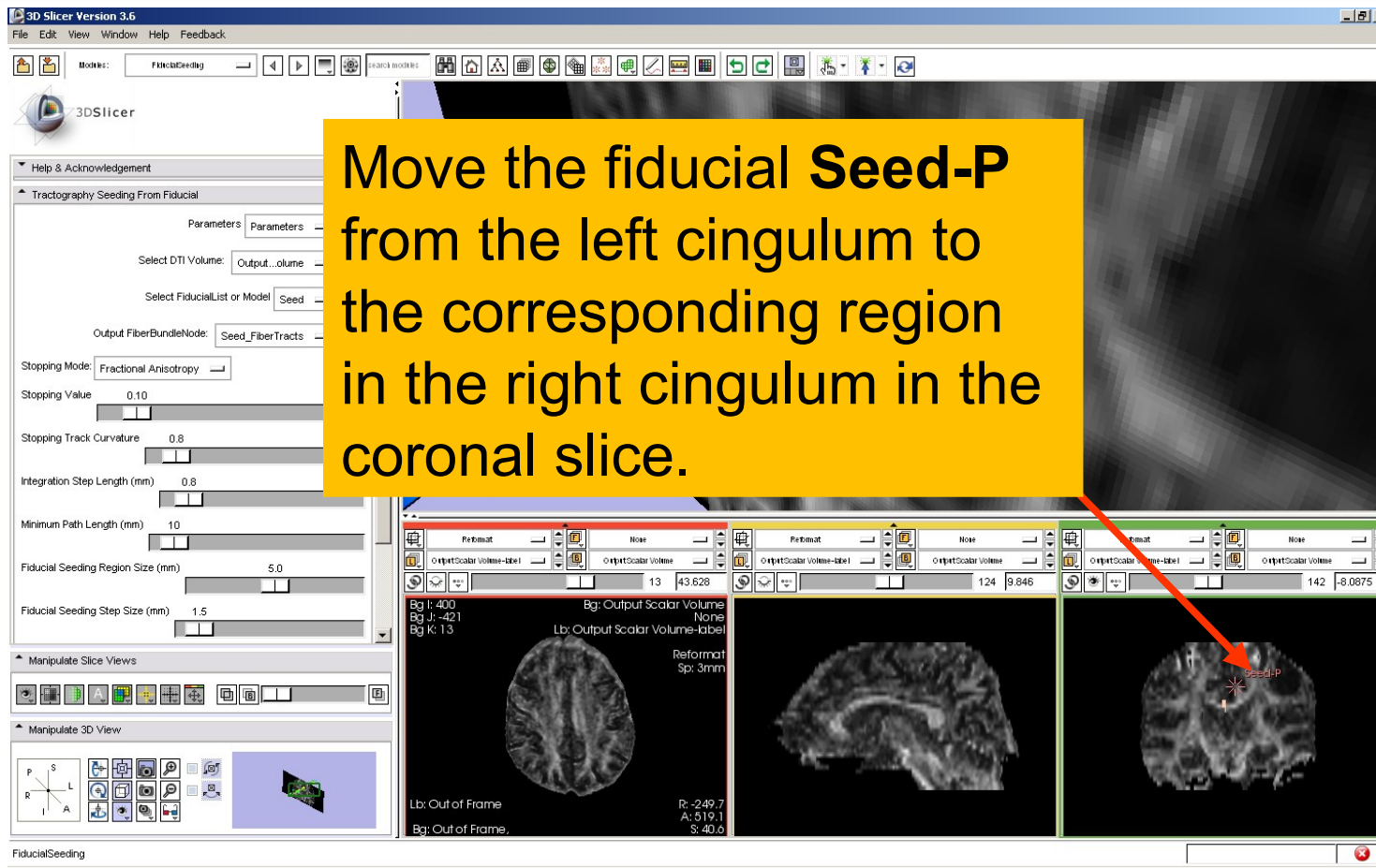
Slicer displays the tracts seeded from the Fiducial Seed-P.

The tracts correspond to the region of the cingulum located above the corpus callosum.

For better visualization, uncheck the visibility box under **Tubes** in the **Fiber Bundles** module (Slide 45).

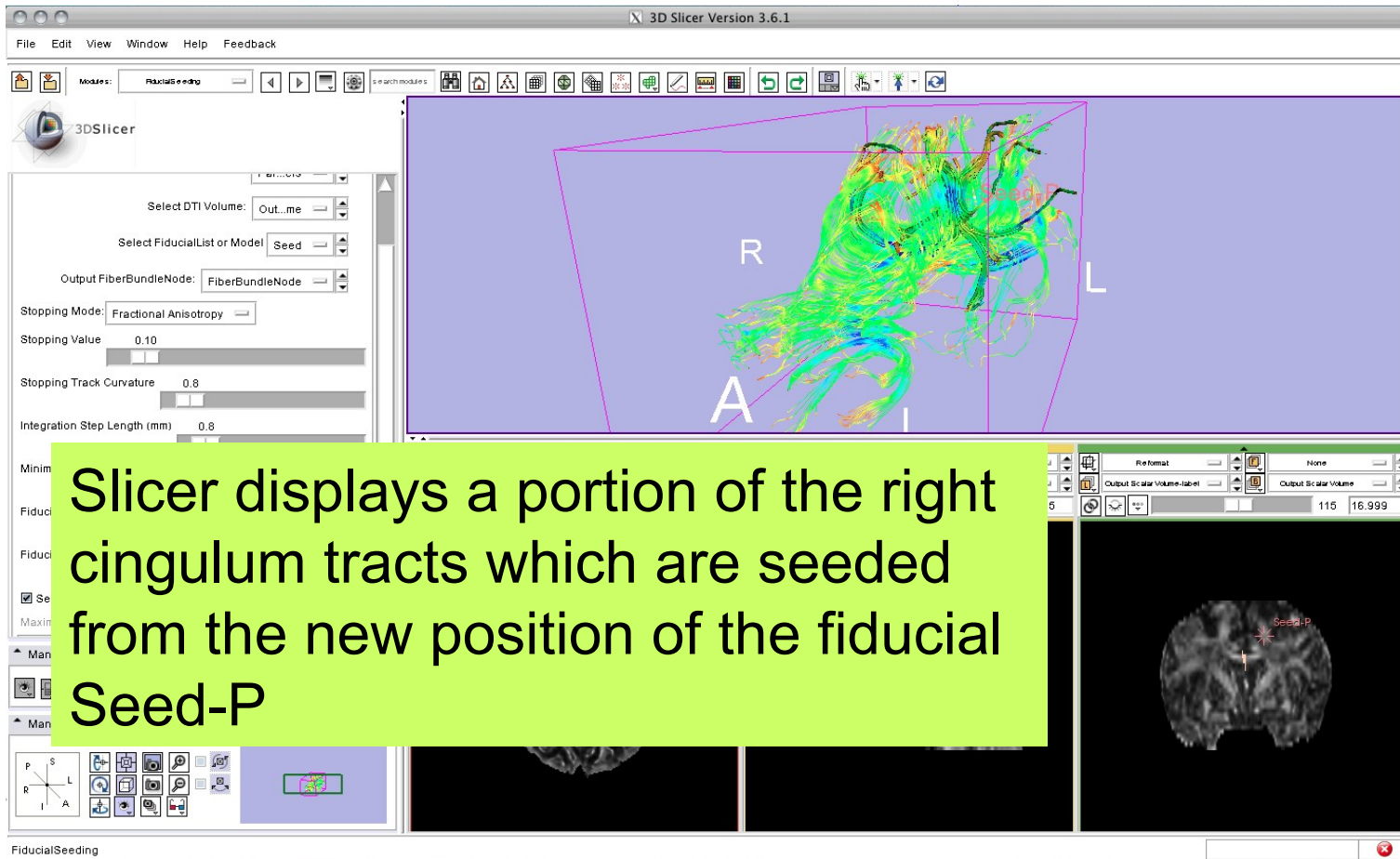


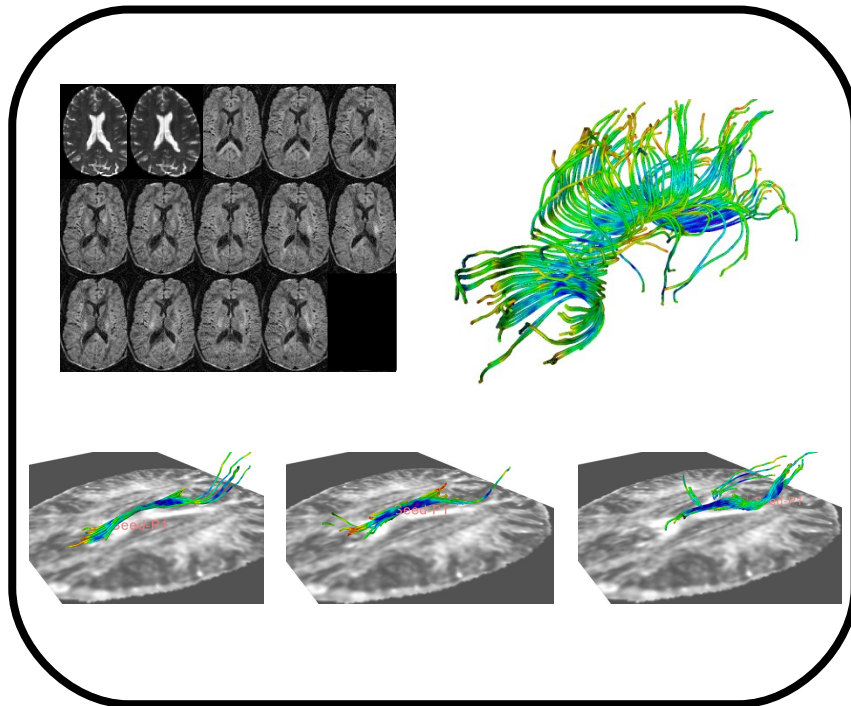
# Fiducial Seeding





# Fiducial Seeding



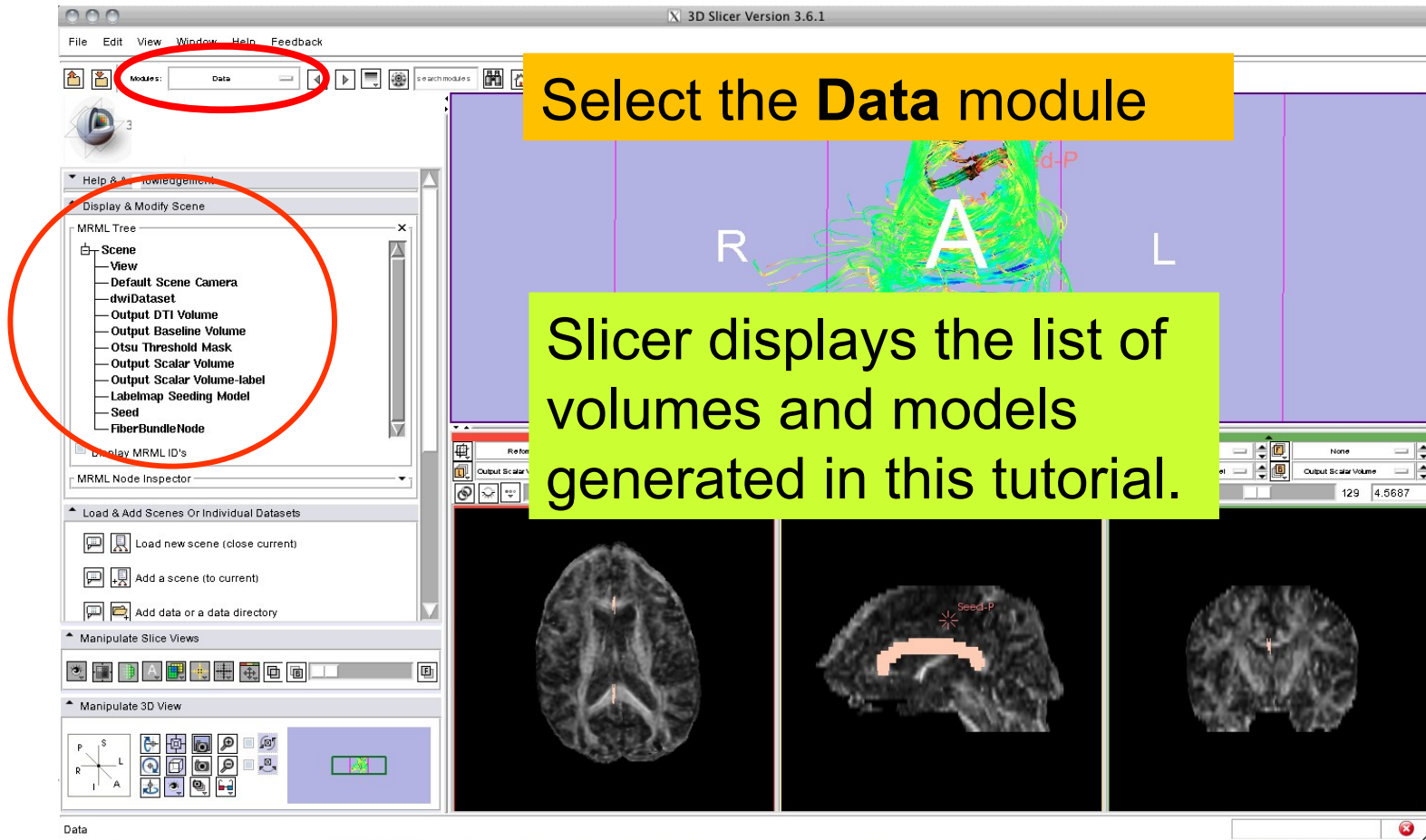


## Part 5:

# Saving a DTI Scene



# DTI Scene







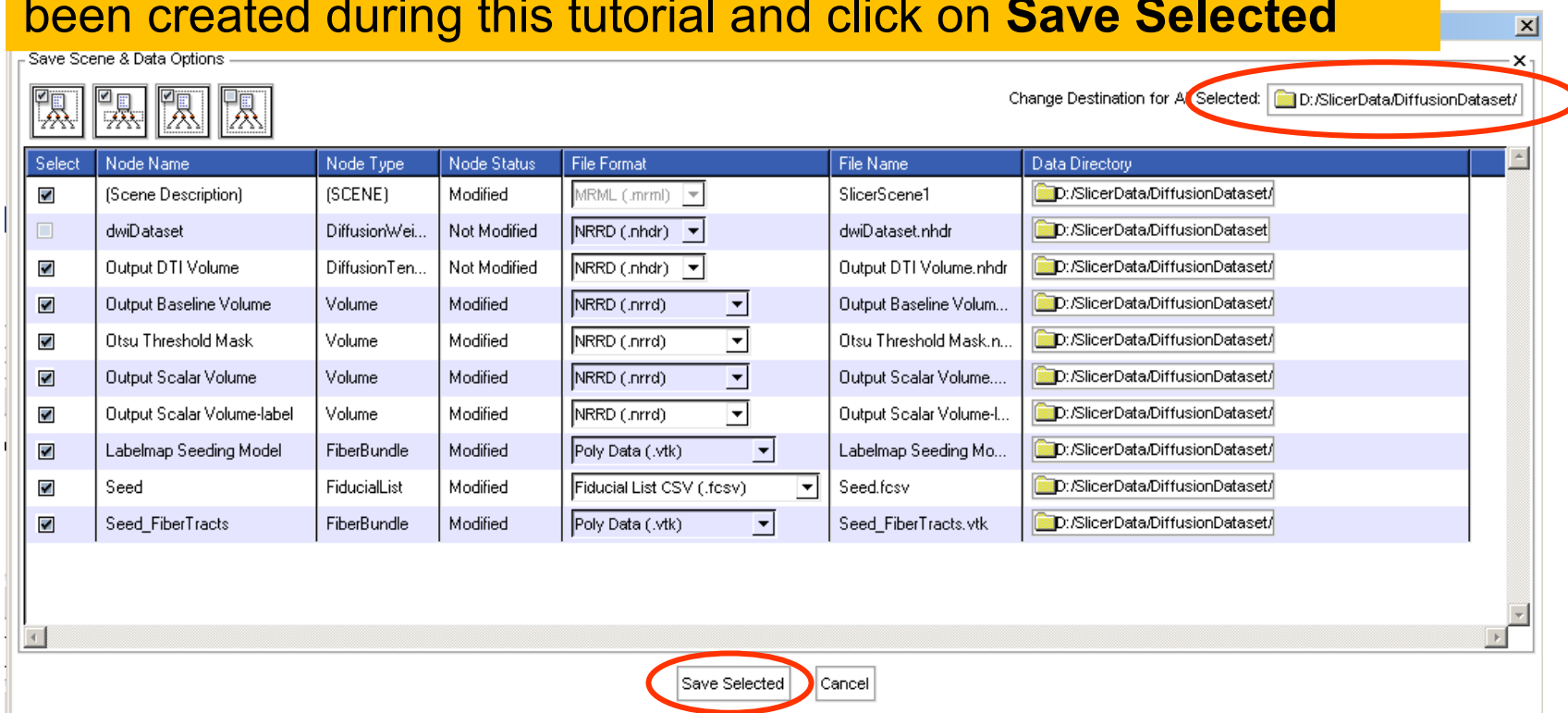
# Saving a DTI Scene





# Saving a DTI Scene

Browse to a directory where you would like to save the data. Once you have selected a directory, select all the files that have been created during this tutorial and click on **Save Selected**



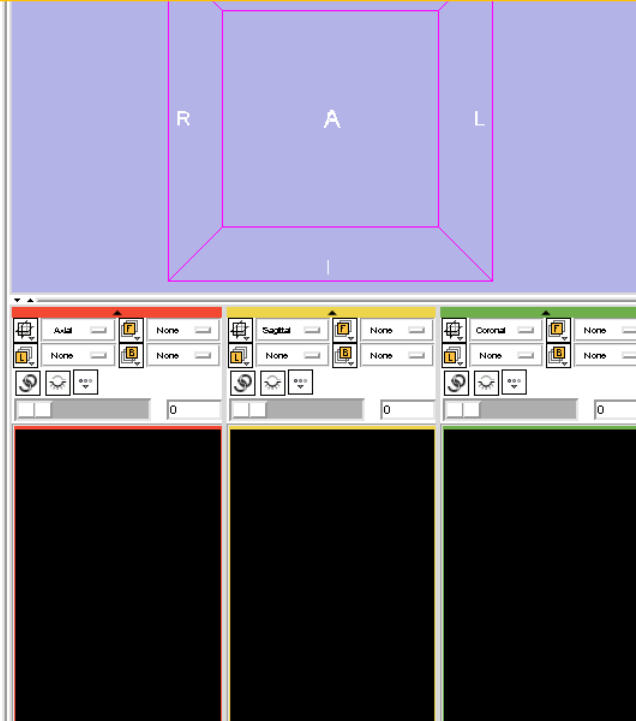




# Saving a DTI Scene



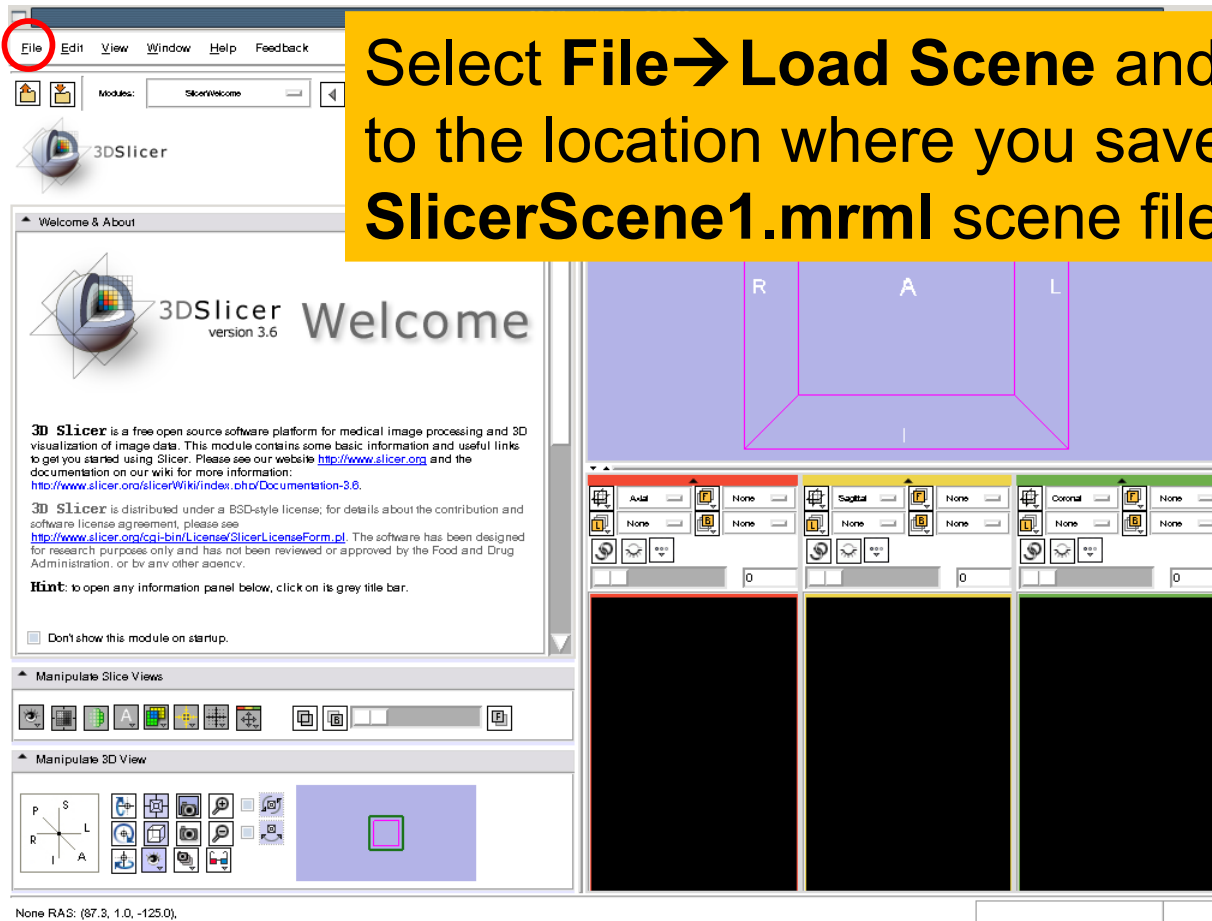
Select **File** → **Close Scene** to close the current DTI Scene



None RAS: (87.3, 1.0, -125.0),

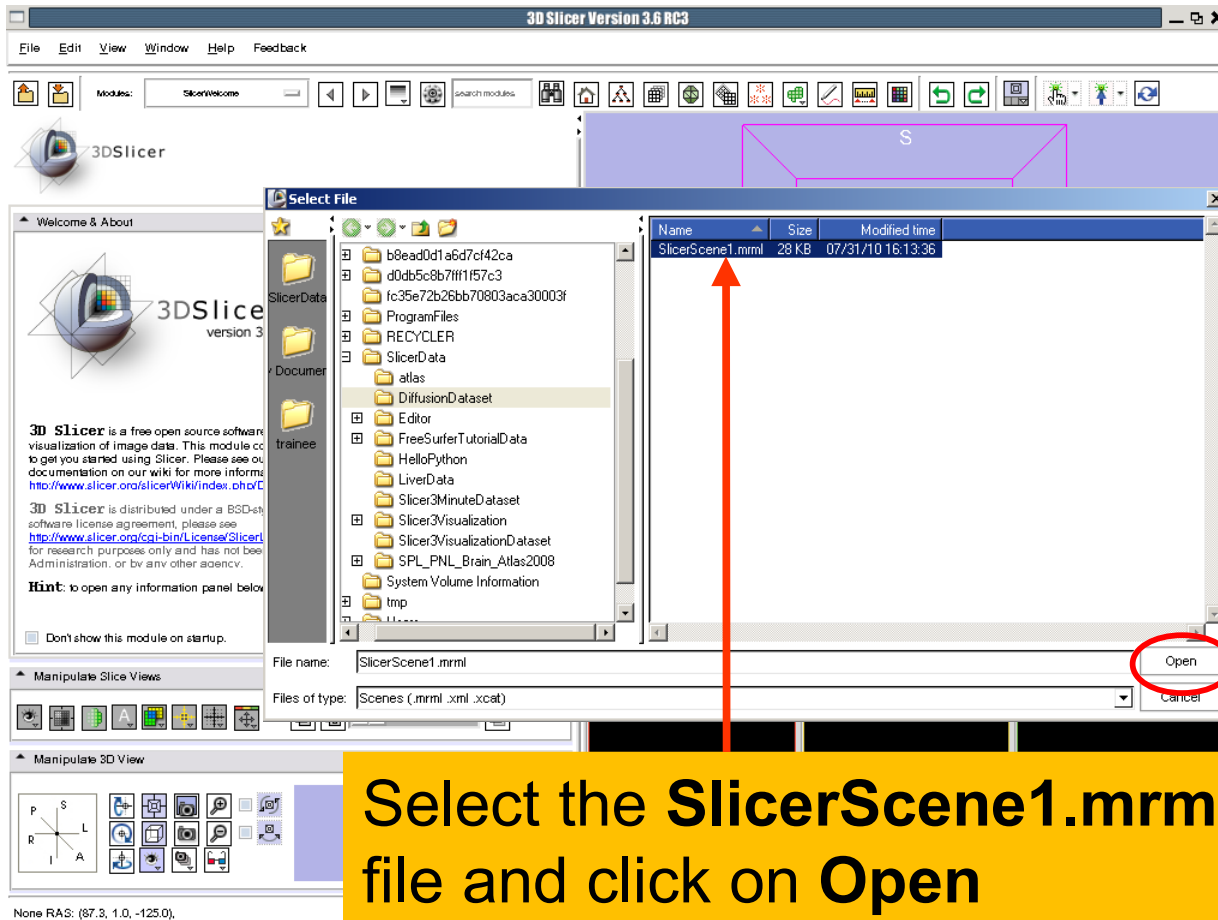


# Loading a DTI Scene



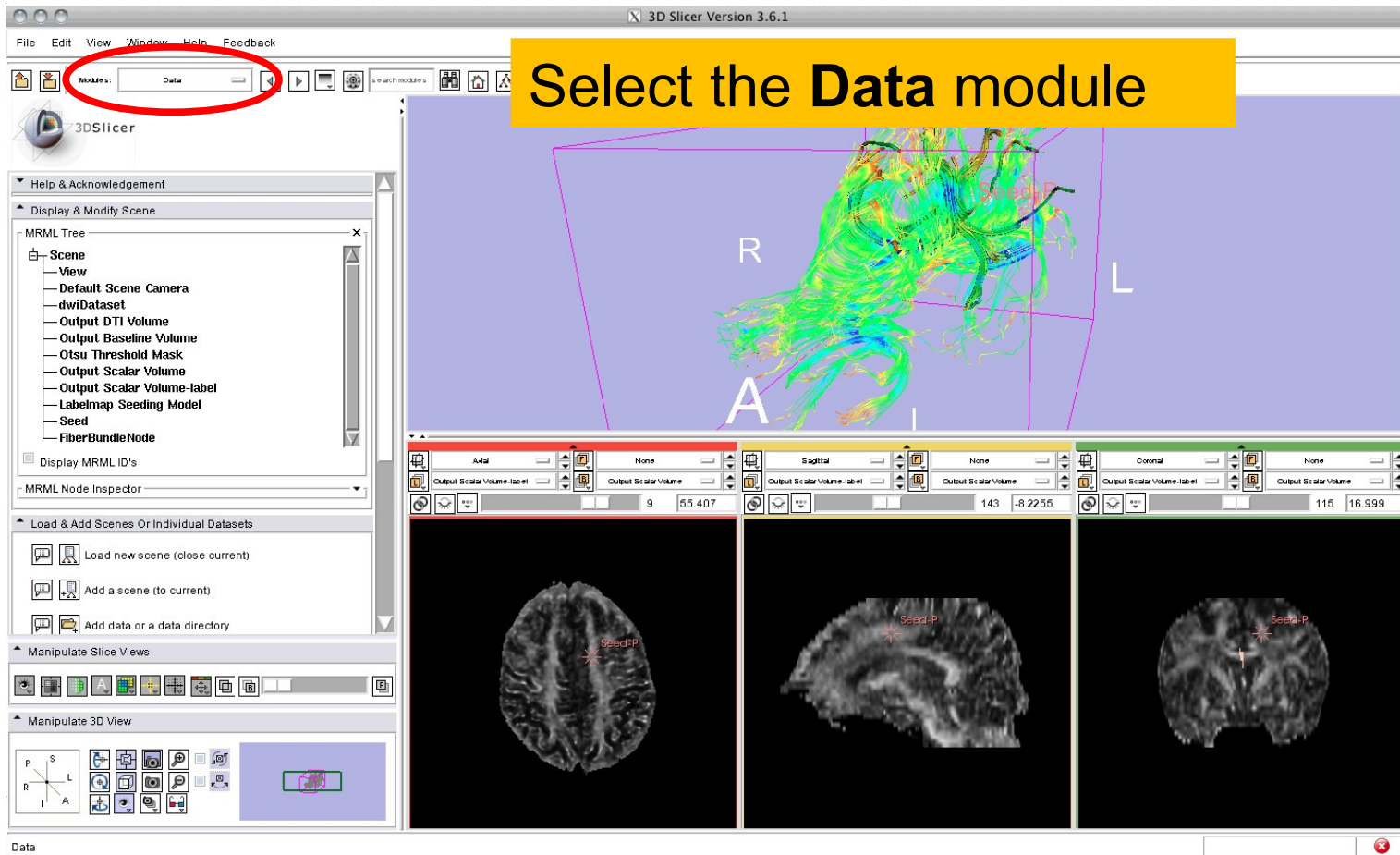


# Loading a DTI Scene





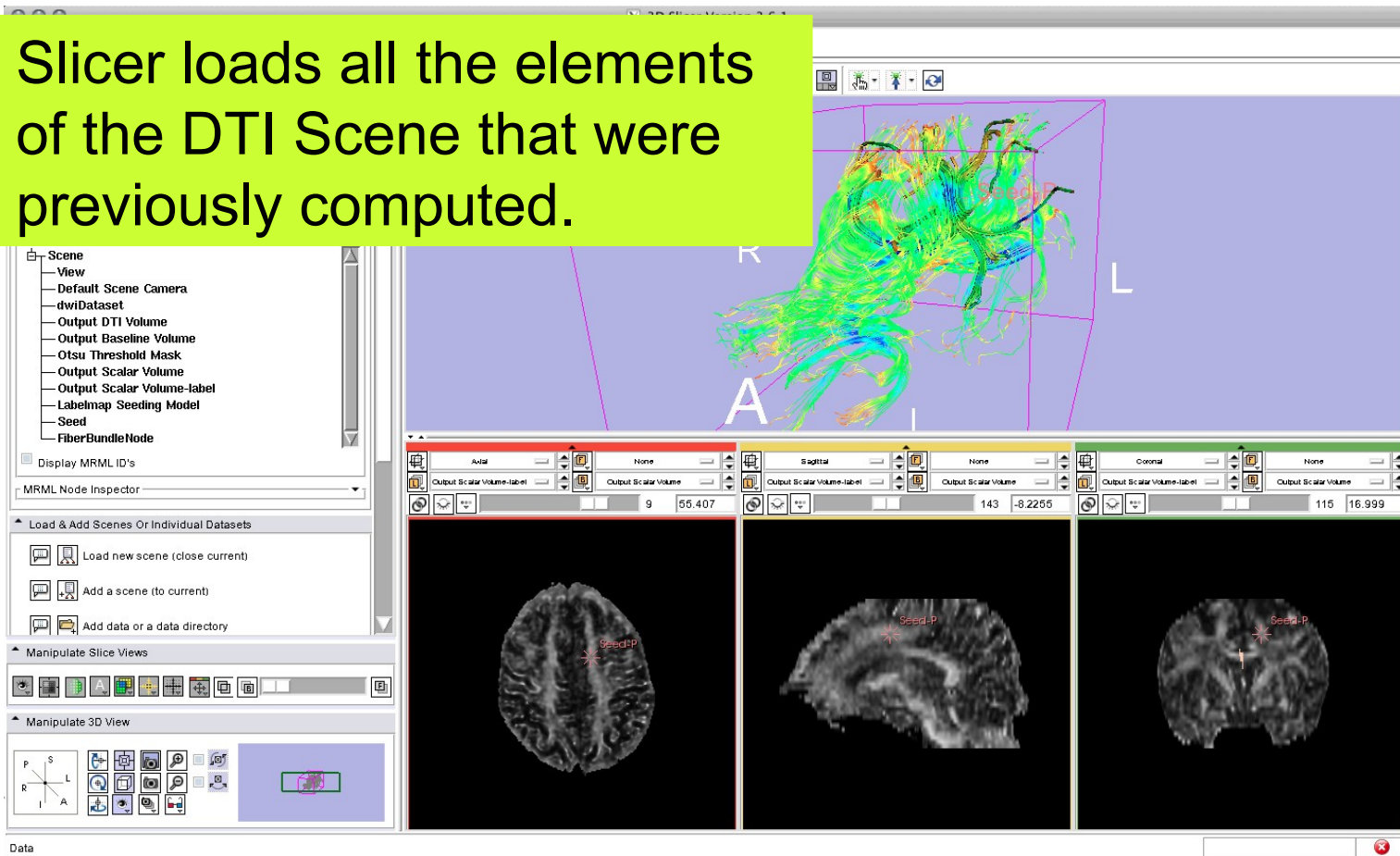
# Loading a DTI Scene





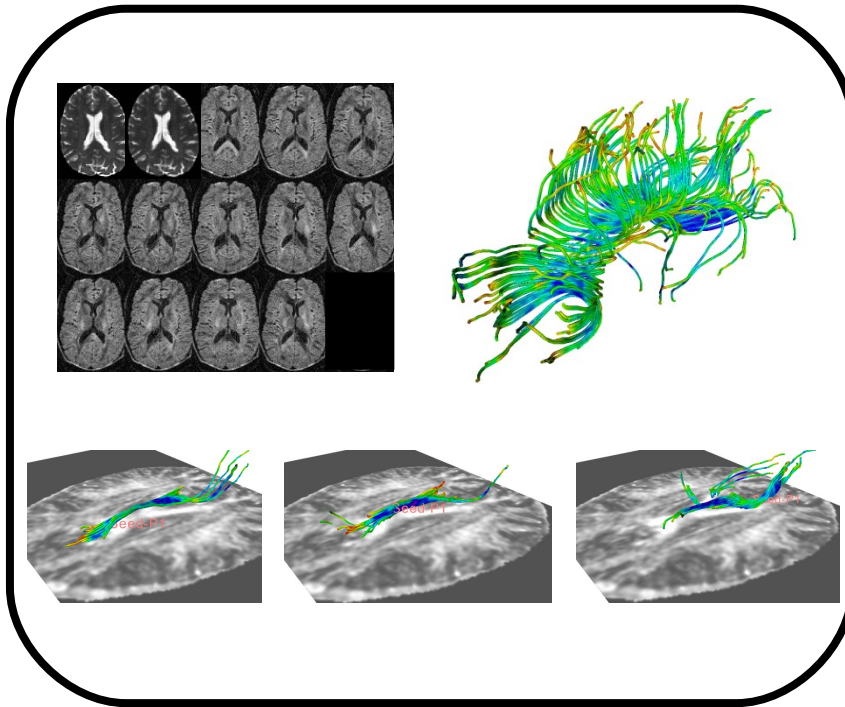
# Loading a DTI Scene

Slicer loads all the elements of the DTI Scene that were previously computed.





# Conclusion



This tutorial guided you through some of the **Diffusion MR** capabilities of the **Slicer3** software for studying the brain white matter pathways.

spujol at bwh.harvard.edu



# Acknowledgments

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